

# Package ‘pls’

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**Title** Partial Least Squares and Principal Component Regression

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**Description** Multivariate regression methods  
Partial Least Squares Regression (PLSR), Principal Component  
Regression (PCR) and Canonical Powered Partial Least Squares (CPPLS).

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**Imports** grDevices, graphics, methods, stats

**Suggests** MASS, parallel, Rmpi, testthat, RUnit

**License** GPL-2

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---

biplot.mvr

*Biplots of PLSR and PCR Models.*

---

## Description

Biplot method for mvr objects.

## Usage

```
## S3 method for class 'mvr'
biplot(
  x,
  comps = 1:2,
  which = c("x", "y", "scores", "loadings"),
  var.axes = FALSE,
```

```

    xlabs,
    ylabs,
    main,
    ...
)

```

### Arguments

x	an mvr object.
comps	integer vector of length two. The components to plot.
which	character. Which matrices to plot. One of "x" (X scores and loadings), "y" (Y scores and loadings), "scores" (X and Y scores) and "loadings" (X and Y loadings).
var.axes	logical. If TRUE, the second set of points have arrows representing them.
xlabs	either a character vector of labels for the first set of points, or FALSE for no labels. If missing, the row names of the first matrix is used as labels.
ylabs	either a character vector of labels for the second set of points, or FALSE for no labels. If missing, the row names of the second matrix is used as labels.
main	character. Title of plot. If missing, a title is constructed by biplot.mvr.
...	Further arguments passed on to biplot.default.

### Details

biplot.mvr can also be called through the mvr plot method by specifying `plottype = "biplot"`.

### Author(s)

Ron Wehrens and Bjørn-Helge Mevik

### See Also

[mvr](#), [plot.mvr](#), [biplot.default](#)

### Examples

```

data(oliveoil)
mod <- pls(sensory ~ chemical, data = oliveoil)
## Not run:
## These are equivalent
biplot(mod)
plot(mod, plottype = "biplot")

## The four combinations of x and y points:
par(mfrow = c(2,2))
biplot(mod, which = "x") # Default
biplot(mod, which = "y")
biplot(mod, which = "scores")
biplot(mod, which = "loadings")

```

```
## End(Not run)
```

---

```
coef.mvr
```

*Extract Information From a Fitted PLSR or PCR Model*

---

## Description

Functions to extract information from mvr objects: Regression coefficients, fitted values, residuals, the model frame, the model matrix, names of the variables and components, and the  $X$  variance explained by the components.

## Usage

```
## S3 method for class 'mvr'
coef(object, ncomp = object$ncomp, comps, intercept = FALSE, ...)

## S3 method for class 'mvr'
fitted(object, ...)

## S3 method for class 'mvr'
residuals(object, ...)

## S3 method for class 'mvr'
model.frame(formula, ...)

## S3 method for class 'mvr'
model.matrix(object, ...)

respnames(object)

prednames(object, intercept = FALSE)

compnames(object, comps, explvar = FALSE, ...)

explvar(object)
```

## Arguments

object, formula	an mvr object. The fitted model.
ncomp, comps	vector of positive integers. The components to include in the coefficients or to extract the names of. See below.
intercept	logical. Whether coefficients for the intercept should be included. Ignored if comps is specified. Defaults to FALSE.
...	other arguments sent to underlying functions. Currently only used for <code>model.frame.mvr</code> and <code>model.matrix.mvr</code> .
explvar	logical. Whether the explained $X$ variance should be appended to the component names.

## Details

These functions are mostly used inside other functions. (Functions `coef.mvr`, `fitted.mvr` and `residuals.mvr` are usually called through their generic functions `coef`, `fitted` and `residuals`, respectively.)

`coef.mvr` is used to extract the regression coefficients of a model, i.e. the  $B$  in  $y = XB$  (for the  $Q$  in  $y = TQ$  where  $T$  is the scores, see [Yloadings](#)). An array of dimension `c(nxvar, nyvar, length(ncomp))` or `c(nxvar, nyvar, length(comps))` is returned.

If `comps` is missing (or is `NULL`), `coef()[, , ncomp[i]]` are the coefficients for models with `ncomp[i]` components, for  $i = 1, \dots, \text{length}(ncomp)$ . Also, if `intercept = TRUE`, the first dimension is `nxvar + 1`, with the intercept coefficients as the first row.

If `comps` is given, however, `coef()[, , comps[i]]` are the coefficients for a model with only the component `comps[i]`, i.e. the contribution of the component `comps[i]` on the regression coefficients.

`fitted.mvr` and `residuals.mvr` return the fitted values and residuals, respectively. If the model was fitted with `na.action = na.exclude` (or after setting the default `na.action` to `"na.exclude"` with [options](#)), the fitted values (or residuals) corresponding to excluded observations are returned as NA; otherwise, they are omitted.

`model.frame.mvr` returns the model frame; i.e. a data frame with all variables necessary to generate the model matrix. See [model.frame](#) for details.

`model.matrix.mvr` returns the (possibly coded) matrix used as  $X$  in the fitting. See [model.matrix](#) for details.

`prednames`, `respnames` and `compnames` extract the names of the  $X$  variables, responses and components, respectively. With `intercept = TRUE` in `prednames`, the name of the intercept variable (i.e. `"(Intercept)"`) is returned as well. `compnames` can also extract component names from score and loading matrices. If `explvar = TRUE` in `compnames`, the explained variance for each component (if available) is appended to the component names. For optimal formatting of the explained variances when not all components are to be used, one should specify the desired components with the argument `comps`.

`explvar` extracts the amount of  $X$  variance (in per cent) explained by each component in the model. It can also handle score and loading matrices returned by [scores](#) and [loadings](#).

## Value

`coef.mvr` returns an array of regression coefficients.

`fitted.mvr` returns an array with fitted values.

`residuals.mvr` returns an array with residuals.

`model.frame.mvr` returns a data frame.

`model.matrix.mvr` returns the  $X$  matrix.

`prednames`, `respnames` and `compnames` return a character vector with the corresponding names.

`explvar` returns a numeric vector with the explained variances, or `NULL` if not available.

## Author(s)

Ron Wehrens and Bjørn-Helge Mevik

**See Also**

[mvr](#), [coef](#), [fitted](#), [residuals](#), [model.frame](#), [model.matrix](#), [na.omit](#)

**Examples**

```
data(yarn)
mod <- pcr(density ~ NIR, data = yarn[yarn$train,], ncomp = 5)
B <- coef(mod, ncomp = 3, intercept = TRUE)
## A manual predict method:
stopifnot(drop(B[1,,] + yarn$NIR[!yarn$train,] %*% B[-1,,]) ==
           drop(predict(mod, ncomp = 3, newdata = yarn[!yarn$train,])))

## Note the difference in formatting:
mod2 <- pcr(density ~ NIR, data = yarn[yarn$train,])
compnames(mod2, explvar = TRUE)[1:3]
compnames(mod2, comps = 1:3, explvar = TRUE)
```

---

coefplot

*Plot Regression Coefficients of PLSR and PCR models*


---

**Description**

Function to plot the regression coefficients of an `mvr` object.

**Usage**

```
coefplot(
  object,
  ncomp = object$ncomp,
  comps,
  intercept = FALSE,
  separate = FALSE,
  se.whiskers = FALSE,
  nCols,
  nRows,
  labels,
  type = "l",
  lty,
  lwd = NULL,
  pch,
  cex = NULL,
  col,
  legendpos,
  xlab = "variable",
  ylab = "regression coefficient",
  main,
  pretty.xlabels = TRUE,
```

```

    xlim,
    ylim,
    ask = nRows * nCols < nPlots && dev.interactive(),
    ...
)

```

## Arguments

object	an mvr object. The fitted model.
ncomp, comps	vector of positive integers. The components to plot. See <a href="#">coef.mvr</a> for details.
intercept	logical. Whether coefficients for the intercept should be plotted. Ignored if comps is specified. Defaults to FALSE. See <a href="#">coef.mvr</a> for details.
separate	logical. If TRUE, coefficients for different model sizes are blotted in separate plots.
se.whiskers	logical. If TRUE, whiskers at plus/minus 1 estimated standard error are added to the plot. This is only available if the model was cross-validated with <code>jackknife = TRUE</code> . Also, in the current implementation, <code>intercept</code> must be FALSE, and <code>separate</code> must be TRUE if <code>length(ncomp) &gt; 1</code> .
nCols, nRows	integer. The number of columns and rows the plots will be laid out in. If not specified, <code>coefplot</code> tries to be intelligent.
labels	optional. Alternative <i>x</i> axis labels. See Details.
type	character. What type of plot to make. Defaults to "l" (lines). Alternative types include "p" (points) and "b" (both). See <a href="#">plot</a> for a complete list of types.
lty	vector of line types (recycled as necessary). Line types can be specified as integers or character strings (see <a href="#">par</a> for the details).
lwd	vector of positive numbers (recycled as necessary), giving the width of the lines.
pch	plot character. A character string or a vector of single characters or integers (recycled as necessary). See <a href="#">points</a> for all alternatives.
cex	numeric vector of character expansion sizes (recycled as necessary) for the plotted symbols.
col	character or integer vector of colors for plotted lines and symbols (recycled as necessary). See <a href="#">par</a> for the details.
legendpos	Legend position. Optional. Ignored if <code>separate</code> is TRUE. If present, a legend is drawn at the given position. The position can be specified symbolically (e.g., <code>legendpos = "topright"</code> ). This requires $\geq 2.1.0$ . Alternatively, the position can be specified explicitly ( <code>legendpos = t(c(x,y))</code> ) or interactively ( <code>legendpos = locator()</code> ). This only works well for plots of single-response models.
xlab, ylab	titles for <i>x</i> and <i>y</i> axes. Typically character strings, but can be expressions (e.g., <code>expression(R^2)</code> ) or lists. See <a href="#">title</a> for details.
main	optional main title for the plot. See Details.
pretty.xlabels	logical. If TRUE, <code>coefplot</code> tries to plot the <i>x</i> labels more nicely. See Details.
xlim, ylim	optional vector of length two, with the <i>x</i> or <i>y</i> limits of the plot.

ask                    logical. Whether to ask the user before each page of a plot.  
 ...                    Further arguments sent to the underlying plot functions.

### Details

coefplot handles multiple responses by making one plot for each response. If `separate` is `TRUE`, separate plots are made for each combination of model size and response. The plots are laid out in a rectangular fashion.

If `legendpos` is given, a legend is drawn at the given position (unless `separate` is `TRUE`).

The argument `labels` can be a vector of labels or one of "names" and "numbers". The labels are used as  $x$  axis labels. If `labels` is "names" or "numbers", the variable names are used as labels, the difference being that with "numbers", the variable names are converted to numbers, if possible. Variable names of the forms "number" or "number text" (where the space is optional), are handled.

The argument `main` can be used to specify the main title of the plot. It is handled in a non-standard way. If there is only one (sub) plot, `main` will be used as the main title of the plot. If there is *more* than one (sub) plot, however, the presence of `main` will produce a corresponding 'global' title on the page. Any graphical parameters, e.g., `cex.main`, supplied to `coefplot` will only affect the 'ordinary' plot titles, not the 'global' one. Its appearance can be changed by setting the parameters with `par`, which will affect *both* titles. (To have different settings for the two titles, one can override the `par` settings with arguments to `coefplot`.)

The argument `pretty.xlabels` is only used when `labels` is specified. If `TRUE` (default), the code tries to use a 'pretty' selection of labels. If `labels` is "numbers", it also uses the numerical values of the labels for horizontal spacing. If one has excluded parts of the spectral region, one might therefore want to use `pretty.xlabels = FALSE`.

When `separate` is `TRUE`, the arguments `lty`, `col`, and `pch` default to their `par()` setting. Otherwise, the default for all of them is `1:nLines`, where `nLines` is the number of model sizes specified, i.e., the length of `ncomp` or `comps`.

The function can also be called through the `mvr` plot method by specifying `plottype = "coefficients"`.

### Note

[legend](#) has many options. If you want greater control over the appearance of the legend, omit the `legendpos` argument and call `legend` manually.

The handling of labels and `pretty.xlabels` is experimental.

### Author(s)

Ron Wehrens and Bjørn-Helge Mevik

### See Also

[mvr](#), [plot.mvr](#), [coef.mvr](#), [plot](#), [legend](#)



**Examples**

```

data(yarn)
mod.nir <- plsr(density ~ NIR, ncomp = 8, data = yarn)
## Not run:
coefplot(mod.nir, ncomp = 1:6)
plot(mod.nir, plottype = "coefficients", ncomp = 1:6) # Equivalent to the previous
## Plot with legend:
coefplot(mod.nir, ncom = 1:6, legendpos = "bottomright")

## End(Not run)

data(oliveoil)
mod.sens <- plsr(sensory ~ chemical, ncomp = 4, data = oliveoil)
## Not run: coefplot(mod.sens, ncomp = 2:4, separate = TRUE)

```

cppls.fit

*CPPLS (Indahl et al.)***Description**

Fits a PLS model using the CPPLS algorithm.

**Usage**

```

cppls.fit(
  X,
  Y,
  ncomp,
  Y.add = NULL,
  center = TRUE,
  stripped = FALSE,
  lower = 0.5,
  upper = 0.5,
  trunc.pow = FALSE,
  weights = NULL,
  ...
)

```

**Arguments**

X	a matrix of observations. NAs and Infs are not allowed.
Y	a vector or matrix of responses. NAs and Infs are not allowed.
ncomp	the number of components to be used in the modelling.
Y.add	a vector or matrix of additional responses containing relevant information about the observations.

center	logical, determines if the $X$ and $Y$ matrices are mean centered or not. Default is to perform mean centering.
stripped	logical. If TRUE the calculations are stripped as much as possible for speed; this is meant for use with cross-validation or simulations when only the coefficients are needed. Defaults to FALSE.
lower	a vector of lower limits for power optimisation. Defaults to 0.5.
upper	a vector of upper limits for power optimisation. Defaults to 0.5.
trunc.pow	logical. If TRUE an experimental alternative power algorithm is used. (Optional)
weights	a vector of individual weights for the observations. (Optional)
...	other arguments. Currently ignored.

### Details

This function should not be called directly, but through the generic functions `cppls` or `mvr` with the argument `method="cppls"`. Canonical Powered PLS (CPPLS) is a generalisation of PLS incorporating discrete and continuous responses (also simultaneously), additional responses, individual weighting of observations and power methodology for sharpening focus on groups of variables. Depending on the input to `cppls` it can produce the following special cases:

- PLS: uni-response continuous  $Y$
- PPLS: uni-response continuous  $Y$ ,  $(\text{lower} \ || \ \text{upper}) \neq 0.5$
- PLS-DA (using correlation maximisation - B/W): dummy-coded discrete response  $Y$
- PPLS-DA: dummy-coded discrete response  $Y$ ,  $(\text{lower} \ || \ \text{upper}) \neq 0.5$
- CPLS: multi-response  $Y$  (continuous, discrete or combination)
- CPPLS: multi-response  $Y$  (continuous, discrete or combination),  $(\text{lower} \ || \ \text{upper}) \neq 0.5$

The name "canonical" comes from canonical correlation analysis which is used when calculating vectors of loading weights, while "powered" refers to a reparameterisation of the vectors of loading weights which can be optimised over a given interval.

### Value

A list containing the following components is returned:

coefficients	an array of regression coefficients for 1, ..., <code>ncomp</code> components. The dimensions of <code>coefficients</code> are <code>c(nvar, npred, ncomp)</code> with <code>nvar</code> the number of $X$ variables and <code>npred</code> the number of variables to be predicted in $Y$ .
scores	a matrix of scores.
loadings	a matrix of loadings.
loading.weights	a matrix of loading weights.
Yscores	a matrix of $Y$ -scores.
Yloadings	a matrix of $Y$ -loadings.
projection	the projection matrix used to convert $X$ to scores.
Xmeans	a vector of means of the $X$ variables.

Ymeans            a vector of means of the Y variables.

fitted.values    an array of fitted values. The dimensions of fitted.values are c(nobj, npred, ncomp) with nobj the number samples and npred the number of Y variables.

residuals        an array of regression residuals. It has the same dimensions as fitted.values.

Xvar             a vector with the amount of X-variance explained by each component.

Xtotvar          total variance in X.

gammas           gamma-values obtained in power optimisation.

canonical.correlations  
                  Canonical correlation values from the calculations of loading weights.

A                 matrix containing vectors of weights a from canonical correlation (cor(Za, Yb)).

smallNorms      vector of indices of explanatory variables of length close to or equal to 0.

If stripped is TRUE, only the components coefficients, Xmeans, Ymeans and gammas are returned.

### Author(s)

Kristian Hovde Liland

### References

Indahl, U. (2005) A twist to partial least squares regression. *Journal of Chemometrics*, **19**, 32–44.

Liland, K.H and Indahl, U.G (2009) Powered partial least squares discriminant analysis, *Journal of Chemometrics*, **23**, 7–18.

Indahl, U.G., Liland, K.H. and Næs, T. (2009) Canonical partial least squares - a unified PLS approach to classification and regression problems. *Journal of Chemometrics*, **23**, 495–504.

### See Also

[mvr](#) [pls](#) [pcr](#) [widekernelpls.fit](#) [simpls.fit](#) [oscorespls.fit](#)

### Examples

```
data(mayonnaise)
# Create dummy response
mayonnaise$dummy <-
  I(model.matrix(~y-1, data.frame(y = factor(mayonnaise$oil.type))))

# Predict CPLS scores for test data
may.cpls <- cppls(dummy ~ NIR, 10, data = mayonnaise, subset = train)
may.test <- predict(may.cpls, newdata = mayonnaise[!mayonnaise$train,], type = "score")

# Predict CPLS scores for test data (experimental used design as additional Y information)
may.cpls.yadd <- cppls(dummy ~ NIR, 10, data = mayonnaise, subset = train, Y.add=design)
may.test.yadd <- predict(may.cpls.yadd, newdata = mayonnaise[!mayonnaise$train,], type = "score")

# Classification by linear discriminant analysis (LDA)
library(MASS)
```

```

error <- matrix(ncol = 10, nrow = 2)
dimnames(error) <- list(Model = c('CPLS', 'CPLS (Y.add)'), ncomp = 1:10)
for (i in 1:10) {
  fitdata1 <- data.frame(oil.type = mayonnaise$oil.type[mayonnaise$train],
                        NIR.score = I(may.cpls$scores[,1:i,drop=FALSE]))
  testdata1 <- data.frame(oil.type = mayonnaise$oil.type[!mayonnaise$train],
                         NIR.score = I(may.test[,1:i,drop=FALSE]))
  error[1,i] <-
    (42 - sum(predict(lda(oil.type ~ NIR.score, data = fitdata1),
                      newdata = testdata1)$class == testdata1$oil.type)) / 42
  fitdata2 <- data.frame(oil.type = mayonnaise$oil.type[mayonnaise$train],
                        NIR.score = I(may.cpls.yadd$scores[,1:i,drop=FALSE]))
  testdata2 <- data.frame(oil.type = mayonnaise$oil.type[!mayonnaise$train],
                         NIR.score = I(may.test.yadd[,1:i,drop=FALSE]))
  error[2,i] <-
    (42 - sum(predict(lda(oil.type ~ NIR.score, data = fitdata2),
                      newdata = testdata2)$class == testdata2$oil.type)) / 42
}
round(error,2)

```

---

crossval

*Cross-validation of PLSR and PCR models*


---

### Description

A “stand alone” cross-validation function for mvr objects.

### Usage

```

crossval(
  object,
  segments = 10,
  segment.type = c("random", "consecutive", "interleaved"),
  length.seg,
  jackknife = FALSE,
  trace = 15,
  ...
)

```

### Arguments

object	an mvr object; the regression to cross-validate.
segments	the number of segments to use, or a list with segments (see below).
segment.type	the type of segments to use. Ignored if segments is a list.
length.seg	Positive integer. The length of the segments to use. If specified, it overrides segments unless segments is a list.
jackknife	logical. Whether jackknifing of regression coefficients should be performed.

trace	if TRUE, tracing is turned on. If numeric, it denotes a time limit (in seconds). If the estimated total time of the cross-validation exceeds this limit, tracing is turned on.
...	additional arguments, sent to the underlying fit function.

## Details

This function performs cross-validation on a model fit by `mvr`. It can handle models such as `pls(y ~ msc(X), ...{ })` or other models where the predictor variables need to be recalculated for each segment. When recalculation is not needed, the result of `crossval(mvr(...{ }))` is identical to `mvr(...{ }, validation = "CV")`, but slower.

Note that to use `crossval`, the data *must* be specified with a `data` argument when fitting object.

If `segments` is a list, the arguments `segment.type` and `length.seg` are ignored. The elements of the list should be integer vectors specifying the indices of the segments. See [cvsegments](#) for details.

Otherwise, segments of type `segment.type` are generated. How many segments to generate is selected by specifying the number of segments in `segments`, or giving the segment length in `length.seg`. If both are specified, `segments` is ignored.

If `jackknife` is TRUE, jackknifed regression coefficients are returned, which can be used for variance estimation ([var.jack](#)) or hypothesis testing ([jack.test](#)).

When tracing is turned on, the segment number is printed for each segment.

By default, the cross-validation will be performed serially. However, it can be done in parallel using functionality in the [parallel](#) package by setting the option `parallel` in [pls.options](#). See [pls.options](#) for the different ways to specify the parallelism. See also Examples below.

## Value

The supplied object is returned, with an additional component `validation`, which is a list with components

<code>method</code>	equals "CV" for cross-validation.
<code>pred</code>	an array with the cross-validated predictions.
<code>coefficients</code>	(only if <code>jackknife</code> is TRUE) an array with the jackknifed regression coefficients. The dimensions correspond to the predictors, responses, number of components, and segments, respectively.
<code>PRESS0</code>	a vector of PRESS values (one for each response variable) for a model with zero components, i.e., only the intercept.
<code>PRESS</code>	a matrix of PRESS values for models with 1, ..., <code>ncomp</code> components. Each row corresponds to one response variable.
<code>adj</code>	a matrix of adjustment values for calculating bias corrected MSE. MSE uses this.
<code>segments</code>	the list of segments used in the cross-validation.
<code>ncomp</code>	the number of components.
<code>gammas</code>	if method <code>cpp1s</code> is used, gamma values for the powers of each CV segment are returned.

**Note**

The PRESS $\hat{\sigma}$  is always cross-validated using leave-one-out cross-validation. This usually makes little difference in practice, but should be fixed for correctness.

The current implementation of the jackknife stores all jackknife-replicates of the regression coefficients, which can be very costly for large matrices. This might change in a future version.

**Author(s)**

Ron Wehrens and Bjørn-Helge Mevik

**References**

Mevik, B.-H., Cederkvist, H. R. (2004) Mean Squared Error of Prediction (MSEP) Estimates for Principal Component Regression (PCR) and Partial Least Squares Regression (PLSR). *Journal of Chemometrics*, **18**(9), 422–429.

**See Also**

[mvr](#) [mvrCv](#) [cvsegments](#) [MSEP](#) [var.jack](#) [jack.test](#)

**Examples**

```
data(yarn)
yarn.pcr <- pcr(density ~ msc(NIR), 6, data = yarn)
yarn.cv <- crossval(yarn.pcr, segments = 10)
## Not run: plot(MSEP(yarn.cv))

## Not run:
## Parallelised cross-validation, using transient cluster:
pls.options(parallel = 4) # use mclapply (not available on Windows)
pls.options(parallel = quote(parallel::makeCluster(4, type = "PSOCK"))) # parLapply
## A new cluster is created and stopped for each cross-validation:
yarn.cv <- crossval(yarn.pcr)
yarn.loocv <- crossval(yarn.pcr, length.seg = 1)

## Parallelised cross-validation, using persistent cluster:
library(parallel)
## This creates the cluster:
pls.options(parallel = makeCluster(4, type = "FORK")) # not available on Windows
pls.options(parallel = makeCluster(4, type = "PSOCK"))
## The cluster can be used several times:
yarn.cv <- crossval(yarn.pcr)
yarn.loocv <- crossval(yarn.pcr, length.seg = 1)
## The cluster should be stopped manually afterwards:
stopCluster(pls.options()$parallel)

## Parallelised cross-validation, using persistent MPI cluster:
## This requires the packages snow and Rmpi to be installed
library(parallel)
## This creates the cluster:
pls.options(parallel = makeCluster(4, type = "MPI"))
```

```
## The cluster can be used several times:
yarn.cv <- crossval(yarn.pcr)
yarn.loocv <- crossval(yarn.pcr, length.seg = 1)
## The cluster should be stopped manually afterwards:
stopCluster(pls.options()$parallel)
## It is good practice to call mpi.exit() or mpi.quit() afterwards:
mpi.exit()

## End(Not run)
```

---

cvsegments

*Generate segments for cross-validation*


---

## Description

The function generates a list of segments for cross-validation. It can generate random, consecutive and interleaved segments, and supports keeping replicates in the same segment.

## Usage

```
cvsegments(
  N,
  k,
  length.seg = ceiling(N/k),
  nrep = 1,
  type = c("random", "consecutive", "interleaved"),
  stratify = NULL
)
```

## Arguments

N	Integer. The number of rows in the data set.
k	Integer. The number of segments to return.
length.seg	Integer. The length of the segments. If given, it overrides k.
nrep	Integer. The number of (consecutive) rows that are replicates of the same object. Replicates will always be kept in the same segment.
type	One of "random", "consecutive" and "interleaved". The type of segments to generate. Default is "random".
stratify	Either a list of indices or an integer vector indicating which stratum each sample (or set of replicates) belongs to (see Details).

## Details

If `length.seg` is specified, it is used to calculate the number of segments to generate. Otherwise `k` must be specified. If  $k * length.seg \neq N$ , the  $k * length.seg - N$  last segments will contain only  $length.seg - 1$  indices.

If `type` is "random", the indices are allocated to segments in random order. If it is "consecutive", the first segment will contain the first  $length.seg$  indices, and so on. If `type` is "interleaved", the first segment will contain the indices  $1, length.seg + 1, 2 * length.seg + 1, \dots, (k - 1) * length.seg + 1$ , and so on.

If `nrep >`, it is assumed that each `nrep` consecutive rows are replicates (repeated measurements) of the same object, and care is taken that replicates are never put in different segments.

Warning: If `k` does not divide `N`, a specified `length.seg` does not divide `N`, or `nrep` does not divide `length.seg`, the number of segments and/or the segment length will be adjusted as needed. Warnings are printed for some of these cases, and one should always inspect the resulting segments to make sure they are as expected.

Stratification of samples is enabled by the `stratify` argument. This is useful if there are subgroups in the data set that should have a proportional representation in the cross-validation segments or if the response is categorical (classification). If `stratify` is combined with `nrep`, `stratify` corresponds to the sets of replicates (see example).

## Value

A list of vectors. Each vector contains the indices for one segment. The attribute "incomplete" contains the number of incomplete segments, and the attribute "type" contains the type of segments.

## Author(s)

Bjørn-Helge Mevik, Ron Wehrens and Kristian Hovde Liland

## Examples

```
## Segments for 10-fold randomised cross-validation:
cvsegments(100, 10)

## Segments with four objects, taken consecutive:
cvsegments(60, length.seg = 4, type = "cons")

## Incomplete segments
segs <- cvsegments(50, length.seg = 3)
attr(segs, "incomplete")

## Leave-one-out cross-validation:
cvsegments(100, 100)
## Leave-one-out with variable/unknown data set size n:
n <- 50
cvsegments(n, length.seg = 1)

## Data set with replicates
cvsegments(100, 25, nrep = 2)
```



```
## Note that rows 1 and 2 are in the same segment, rows 3 and 4 in the
## same segment, and so on.

## Stratification
cvsegments(10, 3, type = "consecutive", stratify = c(rep(1,7), rep(2,3)))
## Note that the last three samples are spread across the segments
## according to the stratification vector.
cvsegments(20, 3, type = "consecutive", nrep = 2, stratify = c(rep(1,7), rep(2,3)))
## Note the length of stratify matching number of replicate sets, not samples.
```

---

 fac2seg

*Factor to Segments*


---

### Description

Factor to Segments

### Usage

```
fac2seg(fac)
```

### Arguments

fac                    A factor where each level represents a segment

### Value

A list of vectors, each vector contains the indices of the elements of the corresponding segment

### Examples

```
fac <- factor(c("a", "b", "a", "b", "c", "c"))
fac2seg(fac)
```

---

 gasoline

*Octane numbers and NIR spectra of gasoline*


---

### Description

A data set with NIR spectra and octane numbers of 60 gasoline samples. The NIR spectra were measured using diffuse reflectance as  $\log(1/R)$  from 900 nm to 1700 nm in 2 nm intervals, giving 401 wavelengths. Many thanks to John H. Kalivas.

**Format**

A data frame with 60 observations on the following 2 variables.

**octane** a numeric vector. The octane number.

**NIR** a matrix with 401 columns. The NIR spectrum.

**Source**

Kalivas, John H. (1997) Two Data Sets of Near Infrared Spectra *Chemometrics and Intelligent Laboratory Systems*, **37**, 255–259.

---

jack.test	<i>Jackknife approximate t tests of regression coefficients</i>
-----------	---

---

**Description**

Performs approximate t tests of regression coefficients based on jackknife variance estimates.

**Usage**

```
jack.test(object, ncomp = object$ncomp, use.mean = TRUE)
```

```
## S3 method for class 'jacktest'
print(x, P.values = TRUE, ...)
```

**Arguments**

object	an mvr object. A cross-validated model fitted with jackknife = TRUE.
ncomp	the number of components to use for estimating the variances
use.mean	logical. If TRUE (default), the mean coefficients are used when estimating the (co)variances; otherwise the coefficients from a model fitted to the entire data set. See <a href="#">var.jack</a> for details.
x	an jacktest object, the result of jack.test.
P.values	logical. Whether to print p values (default).
...	Further arguments sent to the underlying print function <a href="#">printCoefmat</a> .

**Details**

jack.test uses the variance estimates from var.jack to perform t tests of the regression coefficients. The resulting object has a print method, print.jacktest, which uses [printCoefmat](#) for the actual printing.

**Value**

jack.test returns an object of class "jacktest", with components

coefficients	The estimated regression coefficients
sd	The square root of the jackknife variance estimates
tvalues	The $t$ statistics
df	The 'degrees of freedom' used for calculating $p$ values
pvalues	The calculated $p$ values

print.jacktest returns the "jacktest" object (invisibly).

**Warning**

The jackknife variance estimates are known to be biased (see [var.jack](#)). Also, the distribution of the regression coefficient estimates and the jackknife variance estimates are unknown (at least in PLSR/PCR). Consequently, the distribution (and in particular, the degrees of freedom) of the resulting  $t$  statistics is unknown. The present code simply assumes a  $t$  distribution with  $m - 1$  degrees of freedom, where  $m$  is the number of cross-validation segments.

Therefore, the resulting  $p$  values should not be used uncritically, and should perhaps be regarded as mere indicator of (non-)significance.

Finally, also keep in mind that as the number of predictor variables increase, the problem of multiple tests increases correspondingly.

**Author(s)**

Bjørn-Helge Mevik

**References**

Martens H. and Martens M. (2000) Modified Jack-knife Estimation of Parameter Uncertainty in Bilinear Modelling by Partial Least Squares Regression (PLSR). *Food Quality and Preference*, **11**, 5–16.

**See Also**

[var.jack](#), [mvrCv](#)

**Examples**

```
data(oliveoil)
mod <- pcr(sensory ~ chemical, data = oliveoil, validation = "LOO", jackknife = TRUE)
jack.test(mod, ncomp = 2)
```

---

kernelpls.fit	<i>Kernel PLS (Dayal and MacGregor)</i>
---------------	---

---

### Description

Fits a PLSR model with the kernel algorithm.

### Usage

```
kernelpls.fit(X, Y, ncomp, center = TRUE, stripped = FALSE, ...)
```

### Arguments

<code>X</code>	a matrix of observations. NAs and Infs are not allowed.
<code>Y</code>	a vector or matrix of responses. NAs and Infs are not allowed.
<code>ncomp</code>	the number of components to be used in the modelling.
<code>center</code>	logical, determines if the $X$ and $Y$ matrices are mean centered or not. Default is to perform mean centering.
<code>stripped</code>	logical. If TRUE the calculations are stripped as much as possible for speed; this is meant for use with cross-validation or simulations when only the coefficients are needed. Defaults to FALSE.
<code>...</code>	other arguments. Currently ignored.

### Details

This function should not be called directly, but through the generic functions `pls` or `mvr` with the argument `method="kernelpls"` (default). Kernel PLS is particularly efficient when the number of objects is (much) larger than the number of variables. The results are equal to the NIPALS algorithm. Several different forms of kernel PLS have been described in literature, e.g. by De Jong and Ter Braak, and two algorithms by Dayal and MacGregor. This function implements the fastest of the latter, not calculating the crossproduct matrix of  $X$ . In the Dyal & MacGregor paper, this is "algorithm 1".

### Value

A list containing the following components is returned:

<code>coefficients</code>	an array of regression coefficients for 1, ..., <code>ncomp</code> components. The dimensions of <code>coefficients</code> are <code>c(nvar, npred, ncomp)</code> with <code>nvar</code> the number of $X$ variables and <code>npred</code> the number of variables to be predicted in $Y$ .
<code>scores</code>	a matrix of scores.
<code>loadings</code>	a matrix of loadings.
<code>loading.weights</code>	a matrix of loading weights.
<code>Yscores</code>	a matrix of $Y$ -scores.

<code>Yloadings</code>	a matrix of Y-loadings.
<code>projection</code>	the projection matrix used to convert X to scores.
<code>Xmeans</code>	a vector of means of the X variables.
<code>Ymeans</code>	a vector of means of the Y variables.
<code>fitted.values</code>	an array of fitted values. The dimensions of <code>fitted.values</code> are <code>c(nobj, npred, ncomp)</code> with <code>nobj</code> the number samples and <code>npred</code> the number of Y variables.
<code>residuals</code>	an array of regression residuals. It has the same dimensions as <code>fitted.values</code> .
<code>Xvar</code>	a vector with the amount of X-variance explained by each component.
<code>Xtotvar</code>	Total variance in X.

If `stripped` is TRUE, only the components coefficients, `Xmeans` and `Ymeans` are returned.

### Author(s)

Ron Wehrens and Bjørn-Helge Mevik

### References

- de Jong, S. and ter Braak, C. J. F. (1994) Comments on the PLS kernel algorithm. *Journal of Chemometrics*, **8**, 169–174.
- Dayal, B. S. and MacGregor, J. F. (1997) Improved PLS algorithms. *Journal of Chemometrics*, **11**, 73–85.

### See Also

[mvr](#) [pls](#) [cppls](#) [pcr](#) [widekernelpls.fit](#) [simpls.fit](#) [oscorespls.fit](#)

---

mayonnaise

*NIR measurements and oil types of mayonnaise*

---

### Description

Raw NIR measurements (351 wavelengths, 1100-2500 nm in steps of 4 nm) taken on 54 samples of mayonnaise based on six different oil types (soybean, sunflower, canola, olive, corn, and grapeseed). The resulting 54 samples were measured in triplicates, resulting in  $54 \times 3 = 162$  different spectra (120/42 training/test).

### Format

A data frame with 162 observations on the following 4 variables.

**NIR** a matrix with 351 columns

**oil.type** a numeric vector

**design** a matrix with 5 columns

**train** a logical vector

**Source**

Indahl U, Sahni NS, Kirkhus B, Næs T. Multivariate strategies for classification based on NIR-spectra-with application to mayonnaise. *Chemometr. Intell. Lab. Sys.* 1999; 49: 19-31.

---

msc

*Multiplicative Scatter Correction*

---

**Description**

Performs multiplicative scatter/signal correction on a data matrix.

**Usage**

```
msc(X, reference = NULL)

## S3 method for class 'msc'
predict(object, newdata, ...)

## S3 method for class 'msc'
makepredictcall(var, call)
```

**Arguments**

X, newdata	numeric matrices. The data to scatter correct.
reference	numeric vector. Spectre to use as reference. If NULL, the column means of X are used.
object	an object inheriting from class "msc", normally the result of a call to msc with a single matrix argument.
...	other arguments. Currently ignored.
var	A variable.
call	The term in the formula, as a call.

**Details**

makepredictcall.msc is an internal utility function; it is not meant for interactive use. See [makepredictcall](#) for details.

**Value**

Both msc and predict.msc return a multiplicative scatter corrected matrix, with attribute "reference" the vector used as reference spectre. The matrix is given class c("msc", "matrix"). For predict.msc, the "reference" attribute of object is used as reference spectre.

**Author(s)**

Bjørn-Helge Mevik and Ron Wehrens

## References

Martens, H., Næs, T. (1989) *Multivariate calibration*. Chichester: Wiley.

## See Also

[mvr](#), [pcr](#), [plsr](#), [stdize](#)

## Examples

```
data(yarn)
## Direct correction:
Ztrain <- msc(yarn$NIR[yarn$train,])
Ztest <- predict(Ztrain, yarn$NIR[!yarn$train,])

## Used in formula:
mod <- plsr(density ~ msc(NIR), ncomp = 6, data = yarn[yarn$train,])
pred <- predict(mod, newdata = yarn[!yarn$train,]) # Automatically scatter corrected
```

---

mvr

*Partial Least Squares and Principal Component Regression*


---

## Description

Functions to perform partial least squares regression (PLSR), canonical powered partial least squares (CPPLS) or principal component regression (PCR), with a formula interface. Cross-validation can be used. Prediction, model extraction, plot, print and summary methods exist.

## Usage

```
mvr(
  formula,
  ncomp,
  Y.add,
  data,
  subset,
  na.action,
  method = pls.options()$mvralg,
  scale = FALSE,
  center = TRUE,
  validation = c("none", "CV", "LOO"),
  model = TRUE,
  x = FALSE,
  y = FALSE,
  ...
)

plsr(..., method = pls.options()$plsralg)
```

```
pcr(..., method = pls.options()$pcralg)
```

```
cppls(..., Y.add, weights, method = pls.options()$cpplsalg)
```

### Arguments

formula	a model formula. Most of the <code>lm</code> formula constructs are supported. See below.
ncomp	the number of components to include in the model (see below).
Y.add	a vector or matrix of additional responses containing relevant information about the observations. Only used for <code>cppls</code> .
data	an optional data frame with the data to fit the model from.
subset	an optional vector specifying a subset of observations to be used in the fitting process.
na.action	a function which indicates what should happen when the data contain missing values. The default is set by the <code>na.action</code> setting of <code>options</code> , and is <code>na.fail</code> if that is unset. The ‘factory-fresh’ default is <code>na.omit</code> . Another possible value is <code>NULL</code> , no action. Value <code>na.exclude</code> can be useful. See <code>na.omit</code> for other alternatives.
method	the multivariate regression method to be used. If <code>"model.frame"</code> , the model frame is returned.
scale	numeric vector, or logical. If numeric vector, $X$ is scaled by dividing each variable with the corresponding element of <code>scale</code> . If <code>scale</code> is <code>TRUE</code> , $X$ is scaled by dividing each variable by its sample standard deviation. If cross-validation is selected, scaling by the standard deviation is done for every segment.
center	logical, determines if the $X$ and $Y$ matrices are mean centered or not. Default is to perform mean centering.
validation	character. What kind of (internal) validation to use. See below.
model	a logical. If <code>TRUE</code> , the model frame is returned.
x	a logical. If <code>TRUE</code> , the model matrix is returned.
y	a logical. If <code>TRUE</code> , the response is returned.
...	additional optional arguments, passed to the underlying fit functions, and <code>mvrCv</code> . Currently, the fit functions <code>oscorespls.fit</code> and <code>widekernelpls.fit</code> implement these extra arguments: <b>tol:</b> numeric. Tolerance used for determining convergence. <b>maxit:</b> positive integer. The maximal number of iterations used. and <code>cppls.fit</code> implements: <b>lower:</b> a vector of lower limits for power optimisation. <b>upper:</b> a vector of upper limits for power optimisation. <b>trunc.pow:</b> logical. Whether to use an experimental alternative power algorithm. <code>mvrCv</code> implements several arguments; the following are probably the most useful of them:



**segments:** the number of segments to use, or a list with segments.  
**segment.type:** the type of segments to use.  
**length.seg:** Positive integer. The length of the segments to use.  
**jackknife:** logical. Whether to perform jackknifing of regression coefficients.  
 See the functions' documentation for details.

**weights** a vector of individual weights for the observations. Only used for cppls. (Optional)

## Details

The functions fit PLSR, CPPLS or PCR models with 1, ..., ncomp number of components. Multi-response models are fully supported.

The type of model to fit is specified with the method argument. Four PLSR algorithms are available: the kernel algorithm ("kernelpls"), the wide kernel algorithm ("widekernelpls"), SIMPLS ("simpls") and the classical orthogonal scores algorithm ("oscorespls"). One CPPLS algorithm is available ("cppls") providing several extensions to PLS. One PCR algorithm is available: using the singular value decomposition ("svdpc"). If method is "model.frame", the model frame is returned. The functions pcr, pls and cppls are wrappers for mvr, with different values for method.

The formula argument should be a symbolic formula of the form response ~ terms, where response is the name of the response vector or matrix (for multi-response models) and terms is the name of one or more predictor matrices, usually separated by +, e.g., water ~ FTIR or  $y \sim X + Z$ . See [lm](#) for a detailed description. The named variables should exist in the supplied data data frame or in the global environment. Note: Do not use `mvr(mydata$y ~ mydata$X, ...{ })`, instead use `mvr(y ~ X, data = mydata, ...{ })`. Otherwise, `predict.mvr` will not work properly. The chapter 'Statistical models in R' of the manual 'An Introduction to R' distributed with is a good reference on formulas in .

The number of components to fit is specified with the argument ncomp. If this is not supplied, the maximal number of components is used (taking account of any cross-validation).

All implemented algorithms mean-center both predictor and response matrices. This can be turned off by specifying center = FALSE. See Seasholtz and Kowalski for a discussion about centering in PLS regression.

If validation = "CV", cross-validation is performed. The number and type of cross-validation segments are specified with the arguments segments and segment.type. See `mvrCv` for details. If validation = "LOO", leave-one-out cross-validation is performed. It is an error to specify the segments when validation = "LOO" is specified.

By default, the cross-validation will be performed serially. However, it can be done in parallel using functionality in the `parallel` package by setting the option parallel in `pls.options`. See `pls.options` for the different ways to specify the parallelism. See also Examples below.

Note that the cross-validation is optimised for speed, and some generality has been sacrificed. Especially, the model matrix is calculated only once for the complete cross-validation, so models like  $y \sim msc(X)$  will not be properly cross-validated. However, scaling requested by scale = TRUE is properly cross-validated. For proper cross-validation of models where the model matrix must be updated/regenerated for each segment, use the separate function `crossval`.

**Value**

If `method = "model.frame"`, the model frame is returned. Otherwise, an object of class `mvr` is returned. The object contains all components returned by the underlying fit function. In addition, it contains the following components:

<code>validation</code>	if validation was requested, the results of the cross-validation. See <a href="#">mvrCv</a> for details.
<code>fit.time</code>	the elapsed time for the fit. This is used by <a href="#">crossval</a> to decide whether to turn on tracing.
<code>na.action</code>	if observations with missing values were removed, <code>na.action</code> contains a vector with their indices. The class of this vector is used by functions like <code>fitted</code> to decide how to treat the observations.
<code>ncomp</code>	the number of components of the model.
<code>method</code>	the method used to fit the model. See the argument <code>method</code> for possible values.
<code>center</code>	use of centering in the model
<code>scale</code>	if scaling was requested (with <code>scale</code> ), the scaling used.
<code>call</code>	the function call.
<code>terms</code>	the model terms.
<code>model</code>	if <code>model = TRUE</code> , the model frame.
<code>x</code>	if <code>x = TRUE</code> , the model matrix.
<code>y</code>	if <code>y = TRUE</code> , the model response.

**Author(s)**

Ron Wehrens and Bjørn-Helge Mevik

**References**

- Martens, H., Næs, T. (1989) *Multivariate calibration*. Chichester: Wiley.
- Seasholtz, M. B. and Kowalski, B. R. (1992) The effect of mean centering on prediction in multivariate calibration. *Journal of Chemometrics*, **6**(2), 103–111.

**See Also**

[kernelpls.fit](#), [widekernelpls.fit](#), [simpls.fit](#), [oscorespls.fit](#), [cppls.fit](#), [svdpc.fit](#), [mvrCv](#), [crossval](#), [loadings](#), [scores](#), [loading.weights](#), [coef.mvr](#), [predict.mvr](#), [R2](#), [MSEP](#), [RMSEP](#), [plot.mvr](#)

**Examples**

```
data(yarn)
## Default methods:
yarn.pcr <- pcr(density ~ NIR, 6, data = yarn, validation = "CV")
yarn.pls <- pls(density ~ NIR, 6, data = yarn, validation = "CV")
yarn.cppls <- cppls(density ~ NIR, 6, data = yarn, validation = "CV")
```

```

## Alternative methods:
yarn.oscorespls <- mvr(density ~ NIR, 6, data = yarn, validation = "CV",
                      method = "oscorespls")
yarn.simpls <- mvr(density ~ NIR, 6, data = yarn, validation = "CV",
                  method = "simpls")

## Not run:
## Parallelised cross-validation, using transient cluster:
pls.options(parallel = 4) # use mclapply
pls.options(parallel = quote(makeCluster(4, type = "PSOCK"))) # use parLapply
## A new cluster is created and stopped for each cross-validation:
yarn.pls <- plsr(density ~ NIR, 6, data = yarn, validation = "CV")
yarn.pcr <- pcr(density ~ NIR, 6, data = yarn, validation = "CV")

## Parallelised cross-validation, using persistent cluster:
library(parallel)
## This creates the cluster:
pls.options(parallel = makeCluster(4, type = "PSOCK"))
## The cluster can be used several times:
yarn.pls <- plsr(density ~ NIR, 6, data = yarn, validation = "CV")
yarn.pcr <- pcr(density ~ NIR, 6, data = yarn, validation = "CV")
## The cluster should be stopped manually afterwards:
stopCluster(pls.options()$parallel)

## Parallelised cross-validation, using persistent MPI cluster:
## This requires the packages snow and Rmpi to be installed
library(parallel)
## This creates the cluster:
pls.options(parallel = makeCluster(4, type = "MPI"))
## The cluster can be used several times:
yarn.pls <- plsr(density ~ NIR, 6, data = yarn, validation = "CV")
yarn.pcr <- pcr(density ~ NIR, 6, data = yarn, validation = "CV")
## The cluster should be stopped manually afterwards:
stopCluster(pls.options()$parallel)
## It is good practice to call mpi.exit() or mpi.quit() afterwards:
mpi.exit()

## End(Not run)

## Multi-response models:
data(oliveoil)
sens.pcr <- pcr(sensory ~ chemical, ncomp = 4, scale = TRUE, data = oliveoil)
sens.pls <- plsr(sensory ~ chemical, ncomp = 4, scale = TRUE, data = oliveoil)

## Classification
# A classification example utilizing additional response information
# (Y.add) is found in the cppls.fit manual ('See also' above).

```

## Description

Performs the cross-validation calculations for `mvr`.

This function is not meant to be called directly, but through the generic functions `pcr`, `pls`, `cppls` or `mvr` with the argument `validation` set to "CV" or "LOO". All arguments to `mvrCv` can be specified in the generic function call.

If `segments` is a list, the arguments `segment.type` and `length.seg` are ignored. The elements of the list should be integer vectors specifying the indices of the segments. See [cvsegments](#) for details.

Otherwise, segments of type `segment.type` are generated. How many segments to generate is selected by specifying the number of segments in `segments`, or giving the segment length in `length.seg`. If both are specified, `segments` is ignored.

If `jackknife` is TRUE, jackknifed regression coefficients are returned, which can be used for for variance estimation (`var.jack`) or hypothesis testing (`jack.test`).

`X` and `Y` do not need to be centered.

Note that this function cannot be used in situations where  $X$  needs to be recalculated for each segment (except for scaling by the standard deviation), for instance with `msc` or other preprocessing. For such models, use the more general (but slower) function [crossval](#).

Also note that if needed, the function will silently(!) reduce `ncomp` to the maximal number of components that can be cross-validated, which is  $n - l - 1$ , where  $n$  is the number of observations and  $l$  is the length of the longest segment. The (possibly reduced) number of components is returned as the component `ncomp`.

By default, the cross-validation will be performed serially. However, it can be done in parallel using functionality in the [parallel](#) package by setting the option `parallel` in [pls.options](#). See [pls.options](#) for the different ways to specify the parallelism.

## Usage

```
mvrCv(
  X,
  Y,
  ncomp,
  Y.add = NULL,
  weights = NULL,
  method = pls.options()$mvralg,
  scale = FALSE,
  segments = 10,
  segment.type = c("random", "consecutive", "interleaved"),
  length.seg,
  jackknife = FALSE,
  trace = FALSE,
  ...
)
```

## Arguments

`X` a matrix of observations. NAs and Infs are not allowed.

<code>Y</code>	a vector or matrix of responses. NAs and Infs are not allowed.
<code>ncomp</code>	the number of components to be used in the modelling.
<code>Y.add</code>	a vector or matrix of additional responses containing relevant information about the observations. Only used for <code>cpp1s</code> .
<code>weights</code>	a vector of individual weights for the observations. Only used for <code>cpp1s</code> . (Optional)
<code>method</code>	the multivariate regression method to be used.
<code>scale</code>	logical. If TRUE, the learning $X$ data for each segment is scaled by dividing each variable by its sample standard deviation. The prediction data is scaled by the same amount.
<code>segments</code>	the number of segments to use, or a list with segments (see below).
<code>segment.type</code>	the type of segments to use. Ignored if <code>segments</code> is a list.
<code>length.seg</code>	Positive integer. The length of the segments to use. If specified, it overrides <code>segments</code> unless <code>segments</code> is a list.
<code>jackknife</code>	logical. Whether jackknifing of regression coefficients should be performed.
<code>trace</code>	logical; if TRUE, the segment number is printed for each segment.
<code>...</code>	additional arguments, sent to the underlying fit function.

### Value

A list with the following components:

<code>method</code>	equals "CV" for cross-validation.
<code>pred</code>	an array with the cross-validated predictions.
<code>coefficients</code>	(only if <code>jackknife</code> is TRUE) an array with the jackknifed regression coefficients. The dimensions correspond to the predictors, responses, number of components, and segments, respectively.
<code>PRESS0</code>	a vector of PRESS values (one for each response variable) for a model with zero components, i.e., only the intercept.
<code>PRESS</code>	a matrix of PRESS values for models with 1, ..., <code>ncomp</code> components. Each row corresponds to one response variable.
<code>adj</code>	a matrix of adjustment values for calculating bias corrected MSEP. MSEP uses this.
<code>segments</code>	the list of segments used in the cross-validation.
<code>ncomp</code>	the actual number of components used.
<code>gamma</code>	if method <code>cpp1s</code> is used, gamma values for the powers of each CV segment are returned.

### Note

The `PRESS0` is always cross-validated using leave-one-out cross-validation. This usually makes little difference in practice, but should be fixed for correctness.

The current implementation of the `jackknife` stores all `jackknife`-replicates of the regression coefficients, which can be very costly for large matrices. This might change in a future version.

**Author(s)**

Ron Wehrens and Bjørn-Helge Mevik

**References**

Mevik, B.-H., Cederkvist, H. R. (2004) Mean Squared Error of Prediction (MSEP) Estimates for Principal Component Regression (PCR) and Partial Least Squares Regression (PLSR). *Journal of Chemometrics*, **18**(9), 422–429.

**See Also**

[mvr](#) [crossval](#) [cvsegments](#) [MSEP](#) [var](#) [jack](#) [jack.test](#)

**Examples**

```
data(yarn)
yarn.pcr <- pcr(density ~ NIR, 6, data = yarn, validation = "CV", segments = 10)
## Not run: plot(MSEP(yarn.pcr))
```

---

mvrVal

*MSEP, RMSEP and R2 of PLSR and PCR models*


---

**Description**

Functions to estimate the mean squared error of prediction (MSEP), root mean squared error of prediction (RMSEP) and  $R^2$  (A.K.A. coefficient of multiple determination) for fitted PCR and PLSR models. Test-set, cross-validation and calibration-set estimates are implemented.

**Usage**

```
mvrValstats(
  object,
  estimate,
  newdata,
  ncomp = 1:object$ncomp,
  comps,
  intercept = cumulative,
  se = FALSE,
  ...
)

R2(object, ...)

## S3 method for class 'mvr'
R2(
  object,
```

```

    estimate,
    newdata,
    ncomp = 1:object$ncomp,
    comps,
    intercept = cumulative,
    se = FALSE,
    ...
)

MSEP(object, ...)

## S3 method for class 'mvr'
MSEP(
  object,
  estimate,
  newdata,
  ncomp = 1:object$ncomp,
  comps,
  intercept = cumulative,
  se = FALSE,
  ...
)

RMSEP(object, ...)

## S3 method for class 'mvr'
RMSEP(object, ...)

```

### Arguments

<code>object</code>	an <code>mvr</code> object
<code>estimate</code>	a character vector. Which estimators to use. Should be a subset of <code>c("all", "train", "CV", "adjCV", "test")</code> . "adjCV" is only available for (R)MSEP. See below for how the estimators are chosen.
<code>newdata</code>	a data frame with test set data.
<code>ncomp, comps</code>	a vector of positive integers. The components or number of components to use. See below.
<code>intercept</code>	logical. Whether estimates for a model with zero components should be returned as well.
<code>se</code>	logical. Whether estimated standard errors of the estimates should be calculated. Not implemented yet.
<code>...</code>	further arguments sent to underlying functions or (for RMSEP) to MSEP

### Details

RMSEP simply calls MSEP and takes the square root of the estimates. It therefore accepts the same arguments as MSEP.

Several estimators can be used. "train" is the training or calibration data estimate, also called (R)MSEC. For R2, this is the unadjusted  $R^2$ . It is overoptimistic and should not be used for assessing models. "CV" is the cross-validation estimate, and "adjCV" (for RMSEP and MSEP) is the bias-corrected cross-validation estimate. They can only be calculated if the model has been cross-validated. Finally, "test" is the test set estimate, using newdata as test set.

Which estimators to use is decided as follows (see below for mvrValstats). If estimate is not specified, the test set estimate is returned if newdata is specified, otherwise the CV and adjusted CV (for RMSEP and MSEP) estimates if the model has been cross-validated, otherwise the training data estimate. If estimate is "all", all possible estimates are calculated. Otherwise, the specified estimates are calculated.

Several model sizes can also be specified. If comps is missing (or is NULL), length(ncomp) models are used, with ncomp[1] components, ..., ncomp[length(ncomp)] components. Otherwise, a single model with the components comps[1], ..., comps[length(comps)] is used. If intercept is TRUE, a model with zero components is also used (in addition to the above).

The  $R^2$  values returned by "R2" are calculated as  $1 - SSE/SST$ , where  $SST$  is the (corrected) total sum of squares of the response, and  $SSE$  is the sum of squared errors for either the fitted values (i.e., the residual sum of squares), test set predictions or cross-validated predictions (i.e., the *PRESS*). For estimate = "train", this is equivalent to the squared correlation between the fitted values and the response. For estimate = "train", the estimate is often called the prediction  $R^2$ .

mvrValstats is a utility function that calculates the statistics needed by MSEP and R2. It is not intended to be used interactively. It accepts the same arguments as MSEP and R2. However, the estimate argument must be specified explicitly: no partial matching and no automatic choice is made. The function simply calculates the types of estimates it knows, and leaves the other untouched.

## Value

mvrValstats returns a list with components

**SSE** three-dimensional array of SSE values. The first dimension is the different estimators, the second is the response variables and the third is the models.

**SST** matrix of SST values. The first dimension is the different estimators and the second is the response variables.

**nobj** a numeric vector giving the number of objects used for each estimator.

**comps** the components specified, with 0 prepended if intercept is TRUE.

**cumulative** TRUE if comps was NULL or not specified.

The other functions return an object of class "mvrVal", with components

**val** three-dimensional array of estimates. The first dimension is the different estimators, the second is the response variables and the third is the models.

**type** "MSEP", "RMSEP" or "R2".

**comps** the components specified, with 0 prepended if intercept is TRUE.

**cumulative** TRUE if comps was NULL or not specified.

**call** the function call



**Author(s)**

Ron Wehrens and Bjørn-Helge Mevik

**References**

Mevik, B.-H., Cederkvist, H. R. (2004) Mean Squared Error of Prediction (MSEP) Estimates for Principal Component Regression (PCR) and Partial Least Squares Regression (PLSR). *Journal of Chemometrics*, **18**(9), 422–429.

**See Also**

[mvr](#), [crossval](#), [mvrCv](#), [validationplot](#), [plot.mvrVal](#)

**Examples**

```
data(oliveoil)
mod <- pls(sensory ~ chemical, ncomp = 4, data = oliveoil, validation = "L00")
RMSEP(mod)
## Not run: plot(R2(mod))
```

---

oliveoil

*Sensory and physico-chemical data of olive oils*

---

**Description**

A data set with scores on 6 attributes from a sensory panel and measurements of 5 physico-chemical quality parameters on 16 olive oil samples. The first five oils are Greek, the next five are Italian and the last six are Spanish.

**Format**

A data frame with 16 observations on the following 2 variables.

**sensory** a matrix with 6 columns. Scores for attributes ‘yellow’, ‘green’, ‘brown’, ‘glossy’, ‘transp’, and ‘syrup’.

**chemical** a matrix with 5 columns. Measurements of acidity, peroxide, K232, K270, and DK.

**Source**

Massart, D. L., Vandeginste, B. G. M., Buydens, L. M. C., de Jong, S., Lewi, P. J., Smeyers-Verbeke, J. (1998) *Handbook of Chemometrics and Qualimetrics: Part B*. Elsevier. Tables 35.1 and 35.4.

---

oscorespls.fit                      *Orthogonal scores PLSR*

---

## Description

Fits a PLSR model with the orthogonal scores algorithm (aka the NIPALS algorithm).

## Usage

```
oscorespls.fit(
  X,
  Y,
  ncomp,
  center = TRUE,
  stripped = FALSE,
  tol = .Machine$double.eps^0.5,
  maxit = 100,
  ...
)
```

## Arguments

<code>X</code>	a matrix of observations. NAs and Infs are not allowed.
<code>Y</code>	a vector or matrix of responses. NAs and Infs are not allowed.
<code>ncomp</code>	the number of components to be used in the modelling.
<code>center</code>	logical, determines if the $X$ and $Y$ matrices are mean centered or not. Default is to perform mean centering.
<code>stripped</code>	logical. If TRUE the calculations are stripped as much as possible for speed; this is meant for use with cross-validation or simulations when only the coefficients are needed. Defaults to FALSE.
<code>tol</code>	numeric. The tolerance used for determining convergence in multi-response models.
<code>maxit</code>	positive integer. The maximal number of iterations used in the internal Eigen-vector calculation.
<code>...</code>	other arguments. Currently ignored.

## Details

This function should not be called directly, but through the generic functions `pls` or `mvr` with the argument `method="oscorespls"`. It implements the orthogonal scores algorithm, as described in *Martens and Næs (1989)*. This is one of the two “classical” PLSR algorithms, the other being the orthogonal loadings algorithm.

**Value**

A list containing the following components is returned:

<code>coefficients</code>	an array of regression coefficients for 1, ..., ncomp components. The dimensions of <code>coefficients</code> are <code>c(nvar, npred, ncomp)</code> with <code>nvar</code> the number of X variables and <code>npred</code> the number of variables to be predicted in Y.
<code>scores</code>	a matrix of scores.
<code>loadings</code>	a matrix of loadings.
<code>loading.weights</code>	a matrix of loading weights.
<code>Yscores</code>	a matrix of Y-scores.
<code>Yloadings</code>	a matrix of Y-loadings.
<code>projection</code>	the projection matrix used to convert X to scores.
<code>Xmeans</code>	a vector of means of the X variables.
<code>Ymeans</code>	a vector of means of the Y variables.
<code>fitted.values</code>	an array of fitted values. The dimensions of <code>fitted.values</code> are <code>c(nobj, npred, ncomp)</code> with <code>nobj</code> the number samples and <code>npred</code> the number of Y variables.
<code>residuals</code>	an array of regression residuals. It has the same dimensions as <code>fitted.values</code> .
<code>Xvar</code>	a vector with the amount of X-variance explained by each component.
<code>Xtotvar</code>	Total variance in X.

If `stripped` is TRUE, only the components `coefficients`, `Xmeans` and `Ymeans` are returned.

**Author(s)**

Ron Wehrens and Bjørn-Helge Mevik

**References**

Martens, H., Næs, T. (1989) *Multivariate calibration*. Chichester: Wiley.

**See Also**

[mvr](#) [pls](#) [pcr](#) [kernelpls.fit](#) [widekernelpls.fit](#) [simpls.fit](#)

---

`plot.mvr`*Plot Method for MVR objects*

---

## Description

`plot.mvr` plots predictions, coefficients, scores, loadings, biplots, correlation loadings or validation plots (RMSEP curves, etc.).

## Usage

```
## S3 method for class 'mvr'
plot(
  x,
  plottype = c("prediction", "validation", "coefficients", "scores", "loadings",
    "biplot", "correlation"),
  ...
)
```

## Arguments

`x` an object of class `mvr`. The fitted model to plot.

`plottype` character. What kind of plot to plot.

... further arguments, sent to the underlying plot functions.

## Details

The function is simply a wrapper for the underlying plot functions used to make the selected plots. See [predplot.mvr](#), [validationplot](#), [coefplot](#), [scoreplot](#), [loadingplot](#), [biplot.mvr](#) or [corrplot](#) for details. Note that all arguments except `x` and `plottype` must be named.

## Value

`plot.mvr` returns whatever the underlying plot function returns.

## Author(s)

Ron Wehrens and Bjørn-Helge Mevik

## See Also

[mvr](#), [predplot.mvr](#), [validationplot](#), [coefplot](#), [scoreplot](#), [loadingplot](#), [biplot.mvr](#), [corrplot](#)

**Examples**

```

data(yarn)
nir.pcr <- pcr(density ~ NIR, ncomp = 9, data = yarn, validation = "CV")
## Not run:
plot(nir.pcr, ncomp = 5) # Plot of cross-validated predictions
plot(nir.pcr, "scores") # Score plot
plot(nir.pcr, "loadings", comps = 1:3) # The three first loadings
plot(nir.pcr, "coef", ncomp = 5) # Coefficients
plot(nir.pcr, "val") # RMSEP curves
plot(nir.pcr, "val", val.type = "MSEP", estimate = "CV") # CV MSEP

## End(Not run)

```

---

pls.options

Set or return options for the pls package

---

**Description**

A function to set options for the **pls** package, or to return the current options.

**Usage**

```
pls.options(...)
```

**Arguments**

... a single list, a single character vector, or any number of named arguments (*name* = *value*).

**Details**

If called with no arguments, or with an empty list as the single argument, `pls.options` returns the current options.

If called with a character vector as the single argument, a list with the arguments named in the vector are returned.

If called with a non-empty list as the single argument, the list elements should be named, and are treated as named arguments to the function.

Otherwise, `pls.options` should be called with one or more named arguments *name* = *value*. For each argument, the option named *name* will be given the value *value*.

The recognised options are:

**mvrvalg** The fit method to use in `mvr` and `mvrCv`. The value should be one of the allowed methods. Defaults to "kernelpls". Can be overridden with the argument `method` in `mvr` and `mvrCv`.

**pcrvalg** The fit method to use in `pcr`. The value should be one of the allowed methods. Defaults to "svdpc". Can be overridden with the argument `method` in `pcr`.

**plsralg** The fit method to use in `pls`. The value should be one of the allowed methods. Defaults to "kernelpls". Can be overridden with the argument `method` in `pls`.

**cpplsalg** The fit method to use in `cppls`. The value should be one of the allowed methods. Defaults to "cppls". Can be overridden with the argument `method` in `cppls`.

**parallel** Specification of how the cross-validation (CV) in `mvr` should be performed. If the specification is `NULL` (default) or `1`, the CV is done serially, otherwise it is done in parallel using functionality from the `parallel` package.

If it is an integer greater than 1, the CV is done in parallel with the specified number of processes, using `mclapply`.

If it is a cluster object created by `makeCluster`, the CV is done in parallel on that cluster, using `parLapply`. The user should stop the cluster herself when it is no longer needed, using `stopCluster`.

Finally, if the specification is an unevaluated call to `makeCluster`, the call is evaluated, and the CV is done in parallel on the resulting cluster, using `parLapply`. In this case, the cluster will be stopped (with `stopCluster`) after the CV. Thus, in the final case, the cluster is created and destroyed for each CV, just like when using `mclapply`.

**w.tol** The tolerance used for removing values close to 0 in the vectors of loading weights in `cppls`. Defaults to `.Machine$double.eps`.

**X.tol** The tolerance used for removing predictor variables with L1 norms close to 0 in `cppls`. Defaults to  $10^{-12}$ .

### Value

A list with the (possibly changed) options. If any named argument (or list element) was provided, the list is returned invisibly.

### Note

The function is a slight modification of the function `sm.options` from the package `sm`.

### Author(s)

Bjørn-Helge Mevik and Ron Wehrens

### Examples

```
## Return current options:
pls.options()
pls.options("plsralg")
pls.options(c("plsralg", "pcralg"))

## Set options:
pls.options(plsralg = "simpls", mvralg = "simpls")
pls.options(list(plsralg = "simpls", mvralg = "simpls")) # Equivalent
pls.options()

## Restore `factory settings':
pls.options(list(mvralg = "kernelpls", plsralg = "kernelpls", cpplsalg = "cppls",
                pcralg = "svdpc", parallel = NULL,
```

```

                                w.tol = .Machine$double.eps, X.tol = 10^-12))
pls.options()

```

---

predict.mvr

*Predict Method for PLSR and PCR*


---

## Description

Prediction for mvr (PCR, PLSR) models. New responses or scores are predicted using a fitted model and a new matrix of observations.

## Usage

```

## S3 method for class 'mvr'
predict(
  object,
  newdata,
  ncomp = 1:object$ncomp,
  comps,
  type = c("response", "scores"),
  na.action = na.pass,
  ...
)

```

## Arguments

object	an mvr object. The fitted model
newdata	a data frame. The new data. If missing, the training data is used.
ncomp, comps	vector of positive integers. The components to use in the prediction. See below.
type	character. Whether to predict scores or response values
na.action	function determining what should be done with missing values in newdata. The default is to predict NA. See <a href="#">na.omit</a> for alternatives.
...	further arguments. Currently not used

## Details

When type is "response" (default), predicted response values are returned. If comps is missing (or is NULL), predictions for `length(ncomp)` models with `ncomp[1]` components, `ncomp[2]` components, etc., are returned. Otherwise, predictions for a single model with the exact components in comps are returned. (Note that in both cases, the intercept is always included in the predictions. It can be removed by subtracting the `Ymeans` component of the fitted model.)

When type is "scores", predicted score values are returned for the components given in comps. If comps is missing or NULL, `ncomps` is used instead.

It is also possible to supply a matrix instead of a data frame as `newdata`, which is then assumed to be the *X* data matrix. Note that the usual checks for the type of the data are then omitted. Also note that this is *only* possible with `predict`; it will not work in functions like `predplot`, `RMSEP` or `R2`, because they also need the response variable of the new data.

**Value**

When type is "response", a three dimensional array of predicted response values is returned. The dimensions correspond to the observations, the response variables and the model sizes, respectively.

When type is "scores", a score matrix is returned.

**Note**

A warning message like 'newdata' had 10 rows but variable(s) found have 106 rows' means that not all variables were found in the newdata data frame. This (usually) happens if the formula contains terms like yarn\$NIR. Do not use such terms; use the data argument instead. See [mvr](#) for details.

**Author(s)**

Ron Wehrens and Bjørn-Helge Mevik

**See Also**

[mvr](#), [summary.mvr](#), [coef.mvr](#), [plot.mvr](#)

**Examples**

```
data(yarn)
nir.mvr <- mvr(density ~ NIR, ncomp = 5, data = yarn[yarn$train,])

## Predicted responses for models with 1, 2, 3 and 4 components
pred.resp <- predict(nir.mvr, ncomp = 1:4, newdata = yarn[!yarn$train,])

## Predicted responses for a single model with components 1, 2, 3, 4
predict(nir.mvr, comps = 1:4, newdata = yarn[!yarn$train,])

## Predicted scores
predict(nir.mvr, comps = 1:3, type = "scores", newdata = yarn[!yarn$train,])
```

---

predplot

*Prediction Plots*

---

**Description**

Functions to plot predicted values against measured values for a fitted model.



**Usage**

```

predplot(object, ...)

## Default S3 method:
predplot(object, ...)

## S3 method for class 'mvr'
predplot(
  object,
  ncomp = object$ncomp,
  which,
  newdata,
  nCols,
  nRows,
  xlab = "measured",
  ylab = "predicted",
  main,
  ask = nRows * nCols < nPlots && dev.interactive(),
  ...,
  font.main,
  cex.main
)

predplotXy(
  x,
  y,
  line = FALSE,
  labels,
  type = "p",
  main = "Prediction plot",
  xlab = "measured response",
  ylab = "predicted response",
  line.col = par("col"),
  line.lty = NULL,
  line.lwd = NULL,
  ...
)

```

**Arguments**

<code>object</code>	a fitted model.
<code>...</code>	further arguments sent to underlying plot functions.
<code>ncomp</code>	integer vector. The model sizes (numbers of components) to use for prediction.
<code>which</code>	character vector. Which types of predictions to plot. Should be a subset of <code>c("train", "validation", "test")</code> . If not specified, <code>plot.mvr</code> selects test set predictions if <code>newdata</code> is supplied, otherwise cross-validated predictions if the model has been cross-validated, otherwise fitted values from the calibration data.

<code>newdata</code>	data frame. New data to predict.
<code>nCols, nRows</code>	integer. The number of columns and rows the plots will be laid out in. If not specified, <code>plot.mvr</code> tries to be intelligent.
<code>xlab, ylab</code>	titles for $x$ and $y$ axes. Typically character strings, but can be expressions or lists. See <a href="#">title</a> for details.
<code>main</code>	optional main title for the plot. See <a href="#">Details</a> .
<code>ask</code>	logical. Whether to ask the user before each page of a plot.
<code>font.main</code>	font to use for main titles. See <a href="#">par</a> for details. Also see <a href="#">Details</a> below.
<code>cex.main</code>	numeric. The magnification to be used for main titles relative to the current size. Also see <a href="#">Details</a> below.
<code>x</code>	numeric vector. The observed response values.
<code>y</code>	numeric vector. The predicted response values.
<code>line</code>	logical. Whether a target line should be drawn.
<code>labels</code>	optional. Alternative plot labels to use. Either a vector of labels, or "names" or "numbers" to use the row names or row numbers of the data as labels.
<code>type</code>	character. What type of plot to make. Defaults to "p" (points). See <a href="#">plot</a> for a complete list of types. The argument is ignored if <code>labels</code> is specified.
<code>line.col, line.lty, line.lwd</code>	character or numeric. The <code>col</code> , <code>lty</code> and <code>lwd</code> parameters for the target line. See <a href="#">par</a> for details.

## Details

`predplot` is a generic function for plotting predicted versus measured response values, with default and `mvr` methods currently implemented. The default method is very simple, and doesn't handle multiple responses or new data.

The `mvr` method, handles multiple responses, model sizes and types of predictions by making one plot for each combination. It can also be called through the `plot` method for `mvr`, by specifying `plottype = "prediction"` (the default).

The argument `main` can be used to specify the main title of the plot. It is handled in a non-standard way. If there is only one (sub) plot, `main` will be used as the main title of the plot. If there is *more* than one (sub) plot, however, the presence of `main` will produce a corresponding 'global' title on the page. Any graphical parameters, e.g., `cex.main`, supplied to `coefplot` will only affect the 'ordinary' plot titles, not the 'global' one. Its appearance can be changed by setting the parameters with [par](#), which will affect *both* titles (with the exception of `font.main` and `cex.main`, which will only affect the 'global' title when there is more than one plot). (To have different settings for the two titles, one can override the `par` settings with arguments to `predplot`.)

`predplotXy` is an internal function and is not meant for interactive use. It is called by the `predplot` methods, and its arguments, e.g. `line`, can be given in the `predplot` call.

## Value

The functions invisibly return a matrix with the (last) plotted data.

**Note**

The `font.main` and `cex.main` must be (completely) named. This is to avoid that any argument `cex` or `font` matches them.

Tip: If the labels specified with `labels` are too long, they get clipped at the border of the plot region. This can be avoided by supplying the graphical parameter `xpd = TRUE` in the plot call.

**Author(s)**

Ron Wehrens and Bjørn-Helge Mevik

**See Also**

[mvr](#), [plot.mvr](#)

**Examples**

```
data(yarn)
mod <- plsr(density ~ NIR, ncomp = 10, data = yarn[yarn$train,], validation = "CV")
## Not run:
predplot(mod, ncomp = 1:6)
plot(mod, ncomp = 1:6) # Equivalent to the previous
## Both cross-validated and test set predictions:
predplot(mod, ncomp = 4:6, which = c("validation", "test"),
         newdata = yarn[!yarn$train,])

## End(Not run)

data(oliveoil)
mod.sens <- plsr(sensory ~ chemical, ncomp = 4, data = oliveoil)
## Not run: plot(mod.sens, ncomp = 2:4) # Several responses gives several plots
```

---

print.mvr

---

*Summary and Print Methods for PLSR and PCR objects*


---

**Description**

Summary and print methods for `mvr` and `mvrVal` objects.

**Usage**

```
## S3 method for class 'mvr'
print(x, ...)

## S3 method for class 'mvr'
summary(
  object,
  what = c("all", "validation", "training"),
```

```

    digits = 4,
    print.gap = 2,
    ...
)

## S3 method for class 'mvrVal'
print(x, digits = 4, print.gap = 2, ...)

## S3 method for class 'mvrVal'
as.data.frame(x, row.names = NULL, optional = FALSE, shortAlgs = TRUE, ...)

```

**Arguments**

<code>x</code> , object	an <code>mvr</code> object
<code>...</code>	Other arguments sent to underlying methods.
<code>what</code>	one of "all", "validation" or "training"
<code>digits</code>	integer. Minimum number of significant digits in the output. Default is 4.
<code>print.gap</code>	Integer. Gap between columns of the printed tables.
<code>row.names</code>	NULL or a character vector giving the row names for the data frame. Missing values are not allowed.
<code>optional</code>	Not used, only included to match signature of <code>as.data.frame</code> .
<code>shortAlgs</code>	Logical. Shorten algorithm names (default = TRUE).

**Details**

If `what` is "training", the explained variances are given; if it is "validation", the cross-validated RMSEPs (if available) are given; if it is "all", both are given.

**Value**

`print.mvr` and `print.mvrVal` return the object invisibly.

**Author(s)**

Ron Wehrens and Bjørn-Helge Mevik

**See Also**

[mvr](#), [pcr](#), [plsr](#), [RMSEP](#), [MSEP](#)

**Examples**

```

data(yarn)
nir.mvr <- mvr(density ~ NIR, ncomp = 8, validation = "LOO", data = yarn)
nir.mvr
summary(nir.mvr)
RMSEP(nir.mvr)
# Extract MVR validation statistics as data.frame:

```

```
as.data.frame(RMSEP(nir.mvr, estimate = "CV"))
as.data.frame(R2(nir.mvr))
```

---

scoreplot

*Plots of Scores, Loadings and Correlation Loadings*

---

## Description

Functions to make scatter plots of scores or correlation loadings, and scatter or line plots of loadings.

## Usage

```
scoreplot(object, ...)

## Default S3 method:
scoreplot(
  object,
  comps = 1:2,
  labels,
  identify = FALSE,
  type = "p",
  xlab,
  ylab,
  ...
)

## S3 method for class 'scores'
plot(x, ...)

loadingplot(object, ...)

## Default S3 method:
loadingplot(
  object,
  comps = 1:2,
  scatter = FALSE,
  labels,
  identify = FALSE,
  type,
  lty,
  lwd = NULL,
  pch,
  cex = NULL,
  col,
  legendpos,
  xlab,
```

```

    ylab,
    pretty.xlabels = TRUE,
    xlim,
    ...
)

## S3 method for class 'loadings'
plot(x, ...)

corrplot(
  object,
  comps = 1:2,
  labels,
  plotx = TRUE,
  ploty = FALSE,
  radii = c(sqrt(1/2), 1),
  identify = FALSE,
  type = "p",
  xlab,
  ylab,
  col,
  ...
)

```

### Arguments

<code>object</code>	an object. The fitted model.
<code>...</code>	further arguments sent to the underlying plot function(s).
<code>comps</code>	integer vector. The components to plot.
<code>labels</code>	optional. Alternative plot labels or $x$ axis labels. See Details.
<code>identify</code>	logical. Whether to use <code>identify</code> to interactively identify points. See below.
<code>type</code>	character. What type of plot to make. Defaults to "p" (points) for scatter plots and "l" (lines) for line plots. See <a href="#">plot</a> for a complete list of types (not all types are possible/meaningful for all plots).
<code>xlab, ylab</code>	titles for $x$ and $y$ axes. Typically character strings, but can be expressions or lists. See <a href="#">title</a> for details.
<code>x</code>	a scores or loadings object. The scores or loadings to plot.
<code>scatter</code>	logical. Whether the loadings should be plotted as a scatter instead of as lines.
<code>lty</code>	vector of line types (recycled as necessary). Line types can be specified as integers or character strings (see <a href="#">par</a> for the details).
<code>lwd</code>	vector of positive numbers (recycled as necessary), giving the width of the lines.
<code>pch</code>	plot character. A character string or a vector of single characters or integers (recycled as necessary). See <a href="#">points</a> for all alternatives.
<code>cex</code>	numeric vector of character expansion sizes (recycled as necessary) for the plotted symbols.

<code>col</code>	character or integer vector of colors for plotted lines and symbols (recycled as necessary). See <code>par</code> for the details.
<code>legendpos</code>	Legend position. Optional. Ignored if <code>scatter</code> is TRUE. If present, a legend is drawn at the given position. The position can be specified symbolically (e.g., <code>legendpos = "topright"</code> ). This requires $\geq 2.1.0$ . Alternatively, the position can be specified explicitly ( <code>legendpos = t(c(x,y))</code> ) or interactively ( <code>legendpos = locator()</code> ).
<code>pretty.xlabels</code>	logical. If TRUE, <code>loadingplot</code> tries to plot the $x$ labels more nicely. See Details.
<code>xlim</code>	optional vector of length two, with the $x$ limits of the plot.
<code>plotx</code>	logical. Whether to plot the $X$ correlation loadings. Defaults to TRUE.
<code>ploty</code>	logical. Whether to plot the $Y$ correlation loadings. Defaults to FALSE.
<code>radii</code>	numeric vector, giving the radii of the circles drawn in <code>corrplot</code> . The default radii represent 50% and 100% explained variance of the $X$ variables by the chosen components.

## Details

`plot.scores` is simply a wrapper calling `scoreplot`, passing all arguments. Similarly for `plot.loadings`.

`scoreplot` is generic, currently with a default method that works for matrices and any object for which `scores` returns a matrix. The default `scoreplot` method makes one or more scatter plots of the scores, depending on how many components are selected. If one or two components are selected, and `identify` is TRUE, the function `identify` is used to interactively identify points.

Also `loadingplot` is generic, with a default method that works for matrices and any object where `loadings` returns a matrix. If `scatter` is TRUE, the default method works exactly like the default `scoreplot` method. Otherwise, it makes a lineplot of the selected loading vectors, and if `identify` is TRUE, uses `identify` to interactively identify points. Also, if `legendpos` is given, a legend is drawn at the position indicated.

`corrplot` works exactly like the default `scoreplot` method, except that at least two components must be selected. The “correlation loadings”, i.e. the correlations between each variable and the selected components (see References), are plotted as pairwise scatter plots, with concentric circles of radii given by `radii`. Each point corresponds to a variable. The squared distance between the point and origin equals the fraction of the variance of the variable explained by the components in the panel. The default `radii` corresponds to 50% and 100% explained variance. By default, only the correlation loadings of the  $X$  variables are plotted, but if `ploty` is TRUE, also the  $Y$  correlation loadings are plotted.

`scoreplot`, `loadingplot` and `corrplot` can also be called through the `plot` method for `mvr` objects, by specifying `plottype` as “scores”, “loadings” or “correlation”, respectively. See `plot.mvr`.

The argument labels can be a vector of labels or one of “names” and “numbers”.

If a scatter plot is produced (i.e., `scoreplot`, `corrplot`, or `loadingplot` with `scatter = TRUE`), the labels are used instead of plot symbols for the points plotted. If labels is “names” or “numbers”, the row names or row numbers of the matrix (scores, loadings or correlation loadings) are used.

If a line plot is produced (i.e., `loadingplot`), the labels are used as  $x$  axis labels. If labels is “names” or “numbers”, the variable names are used as labels, the difference being that with

"numbers", the variable names are converted to numbers, if possible. Variable names of the forms "number" or "number text" (where the space is optional), are handled.

The argument `pretty.xlabels` is only used when `labels` is specified for a line plot. If `TRUE` (default), the code tries to use a 'pretty' selection of labels. If `labels` is "numbers", it also uses the numerical values of the labels for horizontal spacing. If one has excluded parts of the spectral region, one might therefore want to use `pretty.xlabels = FALSE`.

### Value

The functions return whatever the underlying plot function (or `identify`) returns.

### Note

[legend](#) has many options. If you want greater control over the appearance of the legend, omit the `legendpos` argument and call `legend` manually.

Graphical parameters (such as `pch` and `cex`) can also be used with `scoreplot` and `corrplot`. They are not listed in the argument list simply because they are not handled specifically in the function (unlike in `loadingplot`), but passed directly to the underlying plot functions by `...{}`.

Tip: If the labels specified with `labels` are too long, they get clipped at the border of the plot region. This can be avoided by supplying the graphical parameter `xpd = TRUE` in the plot call.

The handling of `labels` and `pretty.xlabels` in `coefplot` is experimental.

### Author(s)

Ron Wehrens and Bjørn-Helge Mevik

### References

Martens, H., Martens, M. (2000) Modified Jack-knife Estimation of Parameter Uncertainty in Bilinear Modelling by Partial Least Squares Regression (PLSR). *Food Quality and Preference*, **11**(1–2), 5–16.

### See Also

[mvr](#), [plot.mvr](#), [scores](#), [loadings](#), [identify](#), [legend](#)

### Examples

```
data(yarn)
mod <- plsr(density ~ NIR, ncomp = 10, data = yarn)
## These three are equivalent:
## Not run:
scoreplot(mod, comps = 1:5)
plot(scores(mod), comps = 1:5)
plot(mod, plotype = "scores", comps = 1:5)

loadingplot(mod, comps = 1:5)
loadingplot(mod, comps = 1:5, legendpos = "topright") # With legend
loadingplot(mod, comps = 1:5, scatter = TRUE) # Plot as scatterplots
```



```
corrplot(mod, comps = 1:2)
corrplot(mod, comps = 1:3)

## End(Not run)
```

---

scores

*Extract Scores and Loadings from PLSR and PCR Models*

---

## Description

These functions extract score and loading matrices from fitted `mvr` models.

## Usage

```
loadings(object, ...)

## Default S3 method:
loadings(object, ...)

scores(object, ...)

## Default S3 method:
scores(object, ...)

Yscores(object)

loading.weights(object)

Yloadings(object)
```

## Arguments

<code>object</code>	a fitted model to extract from.
<code>...</code>	extra arguments, currently not used.

## Details

All functions extract the indicated matrix from the fitted model, and will work with any object having a suitably named component.

The default scores and loadings methods also handle `prcomp` objects (their scores and loadings components are called `x` and `rotation`, resp.), and add an attribute `"explvar"` with the variance explained by each component, if this is available. (See [`explvar`](#) for details.)

## Value

A matrix with scores or loadings.

**Note**

There is a `loadings` function in package **stats**. It simply returns any element named "loadings". See [loadings](#) for details. The function can be accessed as `stats::loadings(...)`.

**Author(s)**

Ron Wehrens and Bjørn-Helge Mevik

**See Also**

[mvr](#), [coef.mvr](#)

**Examples**

```
data(yarn)
plsmod <- plsr(density ~ NIR, 6, data = yarn)
scores(plsmod)
loadings(plsmod)[,1:4]
```

---

selectNcomp

*Suggestions for the optimal number of components in PCR and PLSR models*

---

**Description**

Choosing the best number of components in PCR and PLSR models is difficult and usually done on the basis of visual inspection of the validation plots. In cases where large numbers of models are built this choice needs to be automated. This function implements two proposals, one based on randomization (permutation) testing, and an approach based on the standard error of the cross-validation residuals.

**Usage**

```
selectNcomp(
  object,
  method = c("randomization", "onesigma"),
  nperm = 999,
  alpha = 0.01,
  ncomp = object$ncomp,
  plot = FALSE,
  ...
)
```

**Arguments**

object	an mvr object. The fitted model. It should contain a validation element.
method	character string, indicating the heuristic to use.
nperm	number of permutations in the "randomization" approach - not used in the "onesigma" approach.
alpha	cutoff for p values in the "randomization" approach - not used in the "onesigma" approach.
ncomp	maximum number of components to consider when determining the global minimum in the cross-validation curve.
plot	whether or not to show a cross-validation plot. The plot for the "randomization" approach shows models that do not differ significantly from the global RMSEP minimum with open circles; the "onesigma" approach shows the one-sigma bands around the RMSEP values. In both cases, the selection is indicated with a blue dashed line.
...	Further plotting arguments, e.g., to add a title to the plot, or to limit the plotting range.

**Details**

In both approaches the results of cross-validation are used, so the model should have been calculated with some form of cross-validation. First, the absolute minimum in the CV curve is determined (considering only the first ncomp components), leading to the reference model. The randomization test approach (Van der Voet, 1994) checks whether the squared prediction errors of models with fewer components are significantly larger than in the reference model. This leads for each model considered to a  $p$  value; the smallest model not significantly worse than the reference model is returned as the selected one.

The approach "onesigma" simply returns the first model where the optimal CV is within one standard error of the absolute optimum (Hastie, Tibshirani and Friedman, 2009). Note that here we simply use the standard deviation of the cross-validation residuals, in line with the procedure used to calculate the error measure itself. Some other packages implementing similar procedures (such as glmnet) calculate an error measure for each validation segment separately and use the average as the final estimate. In such cases the standard error across segments is the relevant measure of spread. For LOO, the two procedures are identical. In other forms of validation, small differences will occur.

**Value**

A number indicating the suggested number of components in the model.

**Author(s)**

Ron Wehrens, Hilko van der Voet and Gerie van der Heijden

**References**

Van der Voet, H. (1994) Comparing the predictive accuracy of models using a simple randomization test. Chemom. Intell. Lab. Syst. 25 (2), 313-323

Hastie, T., Friedman, J. and Tibshirani, R. The Elements of Statistical Learning: data mining, inference, and prediction, Springer (2013), 10th printing with corrections, paragraph 7.10.

### See Also

[mvr](#), [summary.mvr](#)

### Examples

```
data(yarn)
yarn.pls <- plsr(density ~ NIR, data = yarn, scale = TRUE,
               ncomp = 20, validation = "LOO")
selectNcomp(yarn.pls, "onesigma", plot = TRUE, ylim = c(0, 3))
selectNcomp(yarn.pls, "randomization", plot = TRUE)
selectNcomp(yarn.pls, "randomization", plot = TRUE,
           ncomp = 10, ylim = c(0, 3))
```

---

simpls.fit

*Sijmen de Jong's SIMPLS*

---

### Description

Fits a PLSR model with the SIMPLS algorithm.

### Usage

```
simpls.fit(X, Y, ncomp, center = TRUE, stripped = FALSE, ...)
```

### Arguments

X	a matrix of observations. NAs and Infs are not allowed.
Y	a vector or matrix of responses. NAs and Infs are not allowed.
ncomp	the number of components to be used in the modelling.
center	logical, determines if the $X$ and $Y$ matrices are mean centered or not. Default is to perform mean centering.
stripped	logical. If TRUE the calculations are stripped as much as possible for speed; this is meant for use with cross-validation or simulations when only the coefficients are needed. Defaults to FALSE.
...	other arguments. Currently ignored.

### Details

This function should not be called directly, but through the generic functions `plsr` or `mvr` with the argument `method="simpls"`. SIMPLS is much faster than the NIPALS algorithm, especially when the number of  $X$  variables increases, but gives slightly different results in the case of multivariate  $Y$ . SIMPLS truly maximises the covariance criterion. According to de Jong, the standard PLS2 algorithms lie closer to ordinary least-squares regression where a precise fit is sought; SIMPLS lies closer to PCR with stable predictions.

**Value**

A list containing the following components is returned:

<code>coefficients</code>	an array of regression coefficients for 1, ..., <code>ncomp</code> components. The dimensions of <code>coefficients</code> are <code>c(nvar, npred, ncomp)</code> with <code>nvar</code> the number of X variables and <code>npred</code> the number of variables to be predicted in Y.
<code>scores</code>	a matrix of scores.
<code>loadings</code>	a matrix of loadings.
<code>Yscores</code>	a matrix of Y-scores.
<code>Yloadings</code>	a matrix of Y-loadings.
<code>projection</code>	the projection matrix used to convert X to scores.
<code>Xmeans</code>	a vector of means of the X variables.
<code>Ymeans</code>	a vector of means of the Y variables.
<code>fitted.values</code>	an array of fitted values. The dimensions of <code>fitted.values</code> are <code>c(nobj, npred, ncomp)</code> with <code>nobj</code> the number samples and <code>npred</code> the number of Y variables.
<code>residuals</code>	an array of regression residuals. It has the same dimensions as <code>fitted.values</code> .
<code>Xvar</code>	a vector with the amount of X-variance explained by each component.
<code>Xtotvar</code>	Total variance in X.

If `stripped` is TRUE, only the components `coefficients`, `Xmeans` and `Ymeans` are returned.

**Author(s)**

Ron Wehrens and Bjørn-Helge Mevik

**References**

de Jong, S. (1993) SIMPLS: an alternative approach to partial least squares regression. *Chemometrics and Intelligent Laboratory Systems*, **18**, 251–263.

**See Also**

[mvr](#) [pls](#) [pcr](#) [kernelpls.fit](#) [widekernelpls.fit](#) [oscorespls.fit](#)

---

stdize

*Standardization of Data Matrices*

---

**Description**

Performs standardization (centering and scaling) of a data matrix.

**Usage**

```
stdize(x, center = TRUE, scale = TRUE)

## S3 method for class 'stdized'
predict(object, newdata, ...)

## S3 method for class 'stdized'
makepredictcall(var, call)
```

**Arguments**

x, newdata	numeric matrices. The data to standardize.
center	logical value or numeric vector of length equal to the number of columns of x.
scale	logical value or numeric vector of length equal to the number of columns of x.
object	an object inheriting from class "stdized", normally the result of a call to stdize.
...	other arguments. Currently ignored.
var	A variable.
call	The term in the formula, as a call.

**Details**

makepredictcall.stdized is an internal utility function; it is not meant for interactive use. See [makepredictcall](#) for details.

If center is TRUE, x is centered by subtracting the column mean from each column. If center is a numeric vector, it is used in place of the column means.

If scale is TRUE, x is scaled by dividing each column by its sample standard deviation. If scale is a numeric vector, it is used in place of the standard deviations.

**Value**

Both stdize and predict.stdized return a scaled and/or centered matrix, with attributes "stdized:center" and/or "stdized:scale" the vector used for centering and/or scaling. The matrix is given class c("stdized", "matrix").

**Note**

stdize is very similar to [scale](#). The difference is that when scale = TRUE, stdize divides the columns by their standard deviation, while scale uses the root-mean-square of the columns. If center is TRUE, this is equivalent, but in general it is not.

**Author(s)**

Bjørn-Helge Mevik and Ron Wehrens

**See Also**

[mvr](#), [pcr](#), [plsr](#), [msc](#), [scale](#)

**Examples**

```

data(yarn)
## Direct standardization:
Ztrain <- stdize(yarn$NIR[yarn$train,])
Ztest <- predict(Ztrain, yarn$NIR[!yarn$train,])

## Used in formula:
mod <- plsr(density ~ stdize(NIR), ncomp = 6, data = yarn[yarn$train,])
pred <- predict(mod, newdata = yarn[!yarn$train,]) # Automatically standardized

```

svdpc.fit

*Principal Component Regression***Description**

Fits a PCR model using the singular value decomposition.

**Usage**

```
svdpc.fit(X, Y, ncomp, center = TRUE, stripped = FALSE, ...)
```

**Arguments**

<code>X</code>	a matrix of observations. NAs and Infs are not allowed.
<code>Y</code>	a vector or matrix of responses. NAs and Infs are not allowed.
<code>ncomp</code>	the number of components to be used in the modelling.
<code>center</code>	logical, determines if the $X$ and $Y$ matrices are mean centered or not. Default is to perform mean centering.
<code>stripped</code>	logical. If TRUE the calculations are stripped as much as possible for speed; this is meant for use with cross-validation or simulations when only the coefficients are needed. Defaults to FALSE.
<code>...</code>	other arguments. Currently ignored.

**Details**

This function should not be called directly, but through the generic functions `pcr` or `mvr` with the argument `method="svdpc"`. The singular value decomposition is used to calculate the principal components.

**Value**

A list containing the following components is returned:

<code>coefficients</code>	an array of regression coefficients for 1, ..., <code>ncomp</code> components. The dimensions of <code>coefficients</code> are <code>c(nvar, npred, ncomp)</code> with <code>nvar</code> the number of $X$ variables and <code>npred</code> the number of variables to be predicted in $Y$ .
---------------------------	--

scores	a matrix of scores.
loadings	a matrix of loadings.
Yloadings	a matrix of Y-loadings.
projection	the projection matrix used to convert X to scores.
Xmeans	a vector of means of the X variables.
Ymeans	a vector of means of the Y variables.
fitted.values	an array of fitted values. The dimensions of fitted.values are c(nobj, npred, ncomp) with nobj the number samples and npred the number of Y variables.
residuals	an array of regression residuals. It has the same dimensions as fitted.values.
Xvar	a vector with the amount of X-variance explained by each component.
Xtotvar	Total variance in X.

If stripped is TRUE, only the components coefficients, Xmeans and Ymeans are returned.

### Author(s)

Ron Wehrens and Bjørn-Helge Mevik

### References

Martens, H., Næs, T. (1989) *Multivariate calibration*. Chichester: Wiley.

### See Also

[mvr](#) [pls](#) [pcr](#) [cppls](#)

---

validationplot

*Validation Plots*

---

### Description

Functions to plot validation statistics, such as RMSEP or  $R^2$ , as a function of the number of components.

### Usage

```
validationplot(
  object,
  val.type = c("RMSEP", "MSEP", "R2"),
  estimate,
  newdata,
  ncomp,
  comps,
  intercept,
  ...
)
```



```

)

## S3 method for class 'mvrVal'
plot(
  x,
  nCols,
  nRows,
  type = "l",
  lty = 1:nEst,
  lwd = par("lwd"),
  pch = 1:nEst,
  cex = 1,
  col = 1:nEst,
  legendpos,
  xlab = "number of components",
  ylab = x$type,
  main,
  ask = nRows * nCols < nResp && dev.interactive(),
  ...
)

```

### Arguments

object	an <code>mvr</code> object.
val.type	character. What type of validation statistic to plot.
estimate	character. Which estimates of the statistic to calculate. See <a href="#">RMSEP</a> .
newdata	data frame. Optional new data used to calculate statistic.
ncomp, comps	integer vector. The model sizes to compute the statistic for. See <a href="#">RMSEP</a> .
intercept	logical. Whether estimates for a model with zero components should be calculated as well.
...	Further arguments sent to underlying plot functions.
x	an <code>mvrVal</code> object. Usually the result of a <a href="#">RMSEP</a> , <a href="#">MSEP</a> or <a href="#">R2</a> call.
nCols, nRows	integers. The number of columns and rows the plots will be laid out in. If not specified, <code>plot.mvrVal</code> tries to be intelligent.
type	character. What type of plots to create. Defaults to "l" (lines). Alternative types include "p" (points) and "b" (both). See <a href="#">plot</a> for a complete list of types.
lty	vector of line types (recycled as necessary). Line types can be specified as integers or character strings (see <a href="#">par</a> for the details).
lwd	vector of positive numbers (recycled as necessary), giving the width of the lines.
pch	plot character. A character string or a vector of single characters or integers (recycled as necessary). See <a href="#">points</a> for all alternatives.
cex	numeric vector of character expansion sizes (recycled as necessary) for the plotted symbols.

<code>col</code>	character or integer vector of colors for plotted lines and symbols (recycled as necessary). See <a href="#">par</a> for the details.
<code>legendpos</code>	Legend position. Optional. If present, a legend is drawn at the given position. The position can be specified symbolically (e.g., <code>legendpos = "topright"</code> ). This requires $\geq 2.1.0$ . Alternatively, the position can be specified explicitly ( <code>legendpos = t(c(x,y))</code> ) or interactively ( <code>legendpos = locator()</code> ). This only works well for plots of single-response models.
<code>xlab, ylab</code>	titles for $x$ and $y$ axes. Typically character strings, but can be expressions (e.g., <code>expression(R^2)</code> ) or lists. See <a href="#">title</a> for details.
<code>main</code>	optional main title for the plot. See Details.
<code>ask</code>	logical. Whether to ask the user before each page of a plot.

### Details

`validationplot` calls the proper validation function (currently [MSEP](#), [RMSEP](#) or [R2](#)) and plots the results with `plot.mvrVal`. `validationplot` can be called through the `mvr` plot method, by specifying `plottype = "validation"`.

`plot.mvrVal` creates one plot for each response variable in the model, laid out in a rectangle. It uses [matplotlib](#) for performing the actual plotting. If `legendpos` is given, a legend is drawn at the given position.

The argument `main` can be used to specify the main title of the plot. It is handled in a non-standard way. If there is only one (sub) plot, `main` will be used as the main title of the plot. If there is *more* than one (sub) plot, however, the presence of `main` will produce a corresponding ‘global’ title on the page. Any graphical parameters, e.g., `cex.main`, supplied to `coefplot` will only affect the ‘ordinary’ plot titles, not the ‘global’ one. Its appearance can be changed by setting the parameters with [par](#), which will affect *both* titles. (To have different settings for the two titles, one can override the `par` settings with arguments to the plot function.)

### Note

[legend](#) has many options. If you want greater control over the appearance of the legend, omit the `legendpos` argument and call `legend` manually.

### Author(s)

Ron Wehrens and Bjørn-Helge Mevik

### See Also

[mvr](#), [plot.mvr](#), [RMSEP](#), [MSEP](#), [R2](#), [matplotlib](#), [legend](#)

### Examples

```
data(oliveoil)
mod <- plsr(sensory ~ chemical, data = oliveoil, validation = "LOO")
## Not run:
## These three are equivalent:
validationplot(mod, estimate = "all")
```

```

plot(mod, "validation", estimate = "all")
plot(RMSEP(mod, estimate = "all"))
## Plot R2:
plot(mod, "validation", val.type = "R2")
## Plot R2, with a legend:
plot(mod, "validation", val.type = "MSEP", legendpos = "top") # R >= 2.1.0

## End(Not run)

```

---

var.jack

*Jackknife Variance Estimates of Regression Coefficients*


---

### Description

Calculates jackknife variance or covariance estimates of regression coefficients.

The original (Tukey) jackknife variance estimator is defined as  $(g - 1)/g \sum_{i=1}^g (\tilde{\beta}_{-i} - \bar{\beta})^2$ , where  $g$  is the number of segments,  $\tilde{\beta}_{-i}$  is the estimated coefficient when segment  $i$  is left out (called the jackknife replicates), and  $\bar{\beta}$  is the mean of the  $\tilde{\beta}_{-i}$ . The most common case is delete-one jackknife, with  $g = n$ , the number of observations.

This is the definition var.jack uses by default.

However, Martens and Martens (2000) defined the estimator as  $(g - 1)/g \sum_{i=1}^g (\tilde{\beta}_{-i} - \hat{\beta})^2$ , where  $\hat{\beta}$  is the coefficient estimate using the entire data set. I.e., they use the original fitted coefficients instead of the mean of the jackknife replicates. Most (all?) other jackknife implementations for PLSR use this estimator. var.jack can be made to use this definition with use.mean = FALSE. In practice, the difference should be small if the number of observations is sufficiently large. Note, however, that all theoretical results about the jackknife refer to the ‘proper’ definition. (Also note that this option might disappear in a future version.)

### Usage

```
var.jack(object, ncomp = object$ncomp, covariance = FALSE, use.mean = TRUE)
```

### Arguments

object	an mvr object. A cross-validated model fitted with jackknife = TRUE.
ncomp	the number of components to use for estimating the (co)variances
covariance	logical. If TRUE, covariances are calculated; otherwise only variances. The default is FALSE.
use.mean	logical. If TRUE (default), the mean coefficients are used when estimating the (co)variances; otherwise the coefficients from a model fitted to the entire data set. See Details.

**Value**

If covariance is FALSE, an  $p \times q \times c$  array of variance estimates, where  $p$  is the number of predictors,  $q$  is the number of responses, and  $c$  is the number of components.

If covariance is TRUE, an  $pq \times pq \times c$  array of variance-covariance estimates.

**Warning**

Note that the Tukey jackknife variance estimator is not unbiased for the variance of regression coefficients (Hinkley 1977). The bias depends on the  $X$  matrix. For ordinary least squares regression (OLSR), the bias can be calculated, and depends on the number of observations  $n$  and the number of parameters  $k$  in the model. For the common case of an orthogonal design matrix with  $\pm 1$  levels, the delete-one jackknife estimate equals  $(n - 1)/(n - k)$  times the classical variance estimate for the regression coefficients in OLSR. Similar expressions hold for delete-d estimates. Modifications have been proposed to reduce or eliminate the bias for the OLSR case, however, they depend on the number of parameters used in the model. See e.g. Hinkley (1977) or Wu (1986).

Thus, the results of `var.jack` should be used with caution.

**Author(s)**

Bjørn-Helge Mevik

**References**

Tukey J.W. (1958) Bias and Confidence in Not-quite Large Samples. (Abstract of Preliminary Report). *Annals of Mathematical Statistics*, **29**(2), 614.

Martens H. and Martens M. (2000) Modified Jack-knife Estimation of Parameter Uncertainty in Bilinear Modelling by Partial Least Squares Regression (PLSR). *Food Quality and Preference*, **11**, 5–16.

Hinkley D.V. (1977), Jackknifing in Unbalanced Situations. *Technometrics*, **19**(3), 285–292.

Wu C.F.J. (1986) Jackknife, Bootstrap and Other Resampling Methods in Regression Analysis. *The Annals of Statistics*, **14**(4), 1261–1295.

**See Also**

[mvrCv](#), [jack.test](#)

**Examples**

```
data(oliveoil)
mod <- pcr(sensory ~ chemical, data = oliveoil, validation = "LOO",
          jackknife = TRUE)
var.jack(mod, ncomp = 2)
```

---

`vcov.mvr`*Calculate Variance-Covariance Matrix for a Fitted Model Object*

---

## Description

Returns the variance-covariance matrix of the coefficients of a Principal Component Regression.

## Usage

```
## S3 method for class 'mvr'  
vcov(object, ncomp, ...)
```

## Arguments

<code>object</code>	a fitted PCR object of class <code>mvr</code> .
<code>ncomp</code>	number of principal components to estimate <code>vcov</code> for.
<code>...</code>	additional arguments (not used).

## Value

A matrix of estimated covariances between regression coefficients.

## Examples

```
data(yarn)  
yarn.pcr <- pcr(density ~ NIR, 6, data = yarn)  
vc <- vcov(yarn.pcr, 3)  
  
# Standard error of coefficients  
se <- sqrt(diag(vc))  
beta <- coef(yarn.pcr, ncomp = 3)  
  
# Plot regression coefficients with two standard errors shading.  
plot(beta, type = 'l',  
      panel.first = polygon(x = c(1:268, 268:1),  
                            y = c(beta+2*se, rev(beta-2*se)),  
                            col = 'lightblue',  
                            border = NA))
```

---

widekernelpls.fit      *Wide Kernel PLS (Rännar et al.)*

---

## Description

Fits a PLSR model with the wide kernel algorithm.

## Usage

```
widekernelpls.fit(
  X,
  Y,
  ncomp,
  center = TRUE,
  stripped = FALSE,
  tol = .Machine$double.eps^0.5,
  maxit = 100,
  ...
)
```

## Arguments

<code>X</code>	a matrix of observations. NAs and Infs are not allowed.
<code>Y</code>	a vector or matrix of responses. NAs and Infs are not allowed.
<code>ncomp</code>	the number of components to be used in the modelling.
<code>center</code>	logical, determines if the $X$ and $Y$ matrices are mean centered or not. Default is to perform mean centering.
<code>stripped</code>	logical. If TRUE the calculations are stripped as much as possible for speed; this is meant for use with cross-validation or simulations when only the coefficients are needed. Defaults to FALSE.
<code>tol</code>	numeric. The tolerance used for determining convergence in the algorithm.
<code>maxit</code>	positive integer. The maximal number of iterations used in the internal Eigen-vector calculation.
<code>...</code>	other arguments. Currently ignored.

## Details

This function should not be called directly, but through the generic functions `pls` or `mvr` with the argument `method="widekernelpls"`. The wide kernel PLS algorithm is efficient when the number of variables is (much) larger than the number of observations. For very wide  $X$ , for instance  $12 \times 18000$ , it can be twice as fast as `kernelpls.fit` and `simpls.fit`. For other matrices, however, it can be much slower. The results are equal to the results of the NIPALS algorithm.

**Value**

A list containing the following components is returned:

coefficients	an array of regression coefficients for 1, ..., ncomp components. The dimensions of coefficients are c(nvar, npred, ncomp) with nvar the number of X variables and npred the number of variables to be predicted in Y.
scores	a matrix of scores.
loadings	a matrix of loadings.
loading.weights	a matrix of loading weights.
Yscores	a matrix of Y-scores.
Yloadings	a matrix of Y-loadings.
projection	the projection matrix used to convert X to scores.
Xmeans	a vector of means of the X variables.
Ymeans	a vector of means of the Y variables.
fitted.values	an array of fitted values. The dimensions of fitted.values are c(nobj, npred, ncomp) with nobj the number samples and npred the number of Y variables.
residuals	an array of regression residuals. It has the same dimensions as fitted.values.
Xvar	a vector with the amount of X-variance explained by each component.
Xtotvar	Total variance in X.

If stripped is TRUE, only the components coefficients, Xmeans and Ymeans are returned.

**Note**

The current implementation has not undergone extensive testing yet, and should perhaps be regarded as experimental. Specifically, the internal Eigenvector calculation does not always converge in extreme cases where the Eigenvalue is close to zero. However, when it does converge, it always converges to the same results as [kernelpls.fit](#), up to numerical inaccuracies.

The algorithm also has a bit of overhead, so when the number of observations is moderately high, [kernelpls.fit](#) can be faster even if the number of predictors is much higher. The relative speed of the algorithms can also depend greatly on which BLAS and/or LAPACK library is linked against.

**Author(s)**

Bjørn-Helge Mevik

**References**

Rännar, S., Lindgren, F., Geladi, P. and Wold, S. (1994) A PLS Kernel Algorithm for Data Sets with Many Variables and Fewer Objects. Part 1: Theory and Algorithm. *Journal of Chemometrics*, **8**, 111–125.

**See Also**

[mvr.plsr](#) [cppls](#) [pcr](#) [kernelpls.fit](#) [simpls.fit](#) [oscorespls.fit](#)

---

yarn

*NIR spectra and density measurements of PET yarns*

---

**Description**

A training set consisting of 21 NIR spectra of PET yarns, measured at 268 wavelengths, and 21 corresponding densities. A test set of 7 samples is also provided. Many thanks to Erik Swierenga.

**Format**

A data frame with components

**NIR** Numeric matrix of NIR measurements

**density** Numeric vector of densities

**train** Logical vector with TRUE for the training samples and FALSE for the test samples

**Source**

Swierenga H., de Weijer A. P., van Wijk R. J., Buydens L. M. C. (1999) Strategy for constructing robust multivariate calibration models *Chemometrics and Intelligent Laboratory Systems*, **49**(1), 1–17.



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